

ISOLATION AND IDENTIFICATION OF DERMATOPHYTIC FUNGI FROM CATS IN NATHA SATWA NUSANTARA SHELTER, BOGOR, INDONESIA

NICOLE TING QIAN WEN



STUDY PROGRAMME OF VETERINARY MEDICINE SCHOOL OF VETERINARY MEDICINE AND **BIOMEDICAL SCIENCES IPB UNIVERSITY BOGOR** 2025





STATEMENT OF UNDERGRADUATE THESIS, SOURCES OF INFORMATION, AND COPYRIGHT TRANSFER

I hereby declared that my undergraduate thesis entitled "Isolation and Identification of Dermatophytic Fungi from Cats in Natha Satwa Nusantara Shelter, Bogor, Indonesia" is my original work under the supervision of my supervision commission and has not been submitted in any form to other academic institutions. All information derived from other authors, both published and unpublished are mentioned in the references.

I hereby transfer the copyright of my undergraduate thesis to IPB University.

Bogor, July 2025

Nicole Ting Qian Wen B0401211813 . Dilarang mengutip sebagian atau seluruh karya tulis ini tanpa mencantumkan dan menyebutkan sumber

ABSTRACT

NICOLE TING QIAN WEN. Isolation and Identification of Dermatophytic Fungi from Cats in Natha Satwa Nusantara Shelter, Bogor, Indonesia. Supervised by NOVERICKO GINGER BUDIONO and AGUS WIJAYA.

Dermatophytosis is a common zoonotic skin infection caused by dermatophytic fungi in humans and animals including cats. No research has been conducted on dermatophytic fungi in shelter cats within Indonesia. This study determined the prevalence of dermatophytosis in 20 shelter cats from Natha Satwa Nusantara Shelter. All cats were sampled using the MacKenzie toothbrush technique. Additional samples from 14 symptomatic cats were collected via hair plucking (15) and skin scraping (12), total of 27 lesion samples, which were used for direct microscopic examination. Lesions of cats were scored. Wood's lamp test and direct microscopy served as screening tools, while fungal culture on Sabouraud Dextrose Agar (SDA) and Potato Dextrose Agar (PDA), both supplemented with 0.0125% chloramphenicol and 0.04% cycloheximide, act as a diagnostic gold standard. All symptomatic cats showed alopecia (14/14), crusting (8/14), and erythema (3/14). Wood's lamp test was positive in 11/20 cats (55%). All 27 lesion samples revealed fungal elements under microscope. Fungal culture identified Microsporum canis (1/20; 5%) and Trichophyton verrucosum (1/20; 5%), with a total 10% dermatophytosis prevalence. Both positive cats were symptomatic, indicating potential transmission to other animals, humans, and the environment. To reduce the risk of dermatophyte transmission, early detection, isolation of infected animals, environmental disinfection, and routine screening should be implemented as preventive measures.

Keywords: animal shelter, cats, feline dermatophytosis, *M. canis*, *T. verrucosum*



ABSTRAK

NICOLE TING QIAN WEN. Isolasi dan Identifikasi Jamur Dermatofit dari Kucing di Natha Satwa Nusantara Shelter, Bogor, Indonesia. Di bawah bimbingan NOVERICKO GINGER BUDIONO dan AGUS WIJAYA.

Dermatofitosis adalah infeksi kulit zoonosis umum yang disebabkan oleh jamur dermatofit pada manusia dan hewan, termasuk kucing. Belum ada penelitian tentang jamur dermatofit pada kucing di selter Indonesia. Penelitian ini menentukan prevalensi dermatofitosis pada 20 kucing di Natha Satwa Nusantara Selter. Semua kucing diambil sampelnya menggunakan teknik MacKenzie. Sampel tambahan dari 14 kucing bergejala diambil melalui pencabutan rambut (15) dan kerokan kulit (12), total 27 sampel lesi, untuk pemeriksaan mikroskopis langsung. Lesi pada kucing dinilai. Uji Lampu Wood dan mikroskopi langsung berfungsi sebagai alat skrining, dan kultur jamur pada Sabouraud Dextrose Agar (SDA) dan Potato Dextrose Agar (PDA), disuplementasi dengan 0,0125% kloramfenikol dan 0,04% sikloheksimid, digunakan sebagai standar emas diagnostik. Semua kucing bergejala alopesia (14/14), berkerak (8/14), dan eritema (3/14). Uji Lampu Wood positif pada 11/20 kucing (55%). Semua (27 sampel) lesi menunjukkan adanya elemen jamur di bawah mikroskop. Kultur jamur mengidentifikasi Microsporum canis (1/20; 5%) dan Trichophyton verrucosum (1/20; 5%), menghasilkan prevalensi dermatofitosis sebesar 10% (2/20). Kedua kucing yang positif menunjukkan gejala, yang mengindikasikan potensi penularan ke hewan lain, manusia, dan lingkungan. Untuk mengurangi risiko penularan dermatofit, deteksi dini, isolasi hewan yang terinfeksi, desinfeksi lingkungan, dan skrining rutin harus dilakukan sebagai tindakan pencegahan.

Kata kunci: dermatofitosis kucing, kucing, M. canis, selter, T. verrucosum



© Copyright IPB 2025 Copyright is protected by the Law

It is prohibited to cite a part or all this paper without writing or mentioning the source. Citation is only for the purposes of education, research, writing scientific papers, compiling reports, writing criticism, or a review, and the quotation does not harm the interests of IPB.

It is prohibited to publish and reproduce a part or all this paper in any form It is prohibited to publis without permission from IPB.





ISOLATION AND IDENTIFICATION OF DERMATOPHYTIC FUNGI FROM CATS IN NATHA SATWA NUSANTARA SHELTER, BOGOR, INDONESIA

NICOLE TING QIAN WEN

Undergraduate thesis
As one of the requirements to obtain a Bachelor's degree
At the School of Veterinary Medicine and Biomedical
Sciences

STUDY PROGRAMME OF VETERINARY MEDICINE
SCHOOL OF VETERINARY MEDICINE
AND BIOMEDICAL SCIENCES
IPB UNIVERSITY
BOGOR
2025



Examiner in Final Exam:

- 1. Prof. Dr. Dra. R. lis Arifiantini, M.Si.
- 2. Dr. drh. Amaq Fadholly, M.Si.



Title : Isolation and Identification of Dermatophytic Fungi from Cats in Natha

Satwa Nusantara Shelter, Bogor, Indonesia.

Name: Nicole Ting Qian Wen

NIM : B0401211813

Approved by

1st Supervisor:

Dr. drh. Novericko Ginger Budiono, M.Si.

2nd Supervisor: drh. Agus Wijaya M.Sc., Ph.D



Acknowledged by

Head of Veterinary Medicine Undergraduate Study Programme:

Dr. drh. Wahono Esthi Prasetyaningtyas, M.Si NIP. 19800618200642026

Vice Dean of Academic and Student Affairs of School of Veterinary Medicine and Biomedical Sciences:

Prof. drh. Ni Wayan Kurniani Karja, M.P, Ph.D NIP. 196902071996012001



Wahono Esthi Prasetyaningtyas

Date: 12 Jul 2025 09.28.17 WIB Verify at disign.ipb.ac.id





Date of Exam: (10 July 2025)

Date of graduation: 1 4 JUL 2025







ACKNOWLEDGMENT

Assalamualaikum warahmatullahi wabarakatuh. The author expresses praise and gratitude to Allah Subhanahu Wa Ta'ala for granting health and abundant blessings, enabling the completion of this academic work. The research, conducted from September 2024 to April 2025, focuses on the detection of dermatophytes in animals, under the title "Isolation and Identification of Dermatophytic Fungi from Cats in Natha Satwa Nusantara Shelter, Bogor, Indonesia."

The author thanked the Directorate of Research and Innovation of IPB University for providing the funds for this study under the Penelitian Dosen Muda 2024 or Young Lecturers Scheme Fiscal Year 2024 (Number 23434/IT3/PT.01.03/P/B/2024) to Dr. drh. Novericko Ginger Budiono, M.Si. The author thanked the Ethical Committee of the School of Veterinary Medicine and Biomedical Sciences for approving this study's protocol.

The author sincerely thanks research supervisors Dr. drh. Novericko Ginger Budiono, M.Si. and drh. Agus Wijaya, M.Sc., Ph.D., for their invaluable guidance, expertise, and unwavering support throughout this project. Appreciation is also given to drh. Nadira Syahmifariza, the attending veterinarian at Natha Satwa Nusantara Shelter, Bogor, Indonesia, for granting permission to conduct the research, and to Mr. Ismet for his assistance in the laboratory.

The author expresses her deepest gratitude to her family, Mr. Ting Yong Ginn and Mrs. Moh Su Hung, as well as her siblings, Brenda and Frank, for their unwavering support and prayers throughout this academic journey. The author also thanks Ace, Pi Hao, Kai Xuan, Vedha, and all friends who have provided companionship and encouragement during her studies, contributing to the successful completion of this academic work.

Bogor, July 2025

Nicole Ting Qian Wen







TABLE OF CONTENT

LIST OF APPENDICES xx I INTRODUCTION 1.1 1.1. Background 1.1 1.2. Problem 2.1 1.3 Aim 2.2 1.4 Benefit 2.2 II LITERATURE REVIEW 3.2 2.2 Diseases Caused by Fungi in Cat 3.2 2.3 Diagnosis of Fungal Infection in Cat 4.2 2.3.1 Wood Lamp Examination 4.2 2.3.2 Direct Microscopic Examination 4.2 2.3.3 Fungal Culture 5.2 2.4 Shelter Management 5.2 III METHOD 6.6 3.1 Time and Place 6.6 3.2 Study Animals and Materials 6.6 3.3 Methodology 6.7 3.3.1 Sample Collection and Coding 6.6 3.3.2 Anamnesis and Identity Record 6.6 3.3.4 Direct Microscopic Observation 7.7 3.3.5 Fungal Culture 8.8 3.3.6 Macroscopic Observation of SDA Agar Plates 8.3 3.3.7 Microscopic Observation of SDA Agar Plates 8.3 3.4 Data Analysis 9.7 IV RESULTS 1.0 </th <th></th> <th></th> <th></th> <th></th>				
LIST OF APPENDICES	LIS	ST OF TA	ABLE	XX
INTRODUCTION	LIST OF FIGURES			
1.1 Background 1 1.2 Problem 2 1.3 Aim 2 1.4 Benefit 2 II LITERATURE REVIEW 3 2.1 Cats 3 2.2 Diseases Caused by Fungi in Cat 3 2.3 Diagnosis of Fungal Infection in Cat 4 2.3.1 Wood Lamp Examination 4 2.3.2 Direct Microscopic Examination 4 2.3.2 Direct Microscopic Examination 5 2.4 Shelter Management 6 3.1 Time and Place 6 3.2 Study Animals and Materials 6 3.3 Methodology 6 3.3.1 Sample Collection and Coding 6 3.3.2 Anamnesis and Identity Record 6 3.3.3 Sample Collection 6 3.3.4 Direct Microscopic Observation 7 3.3.5 Fungal Culture 8 3.3.6 Macroscopic Observation of SDA Agar Plates 8 3.4 Data Analysis 9 <	LIS	T OF A	PPENDICES	XX
2.1 Cats 2.2 Diseases Caused by Fungi in Cat 3 2.3 Diagnosis of Fungal Infection in Cat 4 2.3.1 Wood Lamp Examination 4 2.3.2 Direct Microscopic Examination 4 2.3.3 Fungal Culture 5 2.4 Shelter Management 6 3.1 Time and Place 6 3.2 Study Animals and Materials 6 3.3 Methodology 6 3.3.1 Sample Collection and Coding 6 3.3.2 Anamnesis and Identity Record 6 3.3.4 Direct Microscopic Observation 7 3.3.5 Fungal Culture 8 3.3.6 Macroscopic Observation of SDA Agar Plates 8 3.3.7 Microscopic Observation of SDA Agar Plates 8 3.4 Data Analysis 9 IV RESULTS 10 4.1 Cats 10 4.2 Direct Examination 13 4.3 Identification of Isolated Dermatophytes 14 4.4 Prevalence of Dermatophytes 20 </td <td>I</td> <td>1.1 1.2 1.3</td> <td>Background Problem Aim</td> <td>1 1 2 2 2 2</td>	I	1.1 1.2 1.3	Background Problem Aim	1 1 2 2 2 2
III METHOD 3.1 Time and Place 3.2 Study Animals and Materials 3.3 Methodology 3.3.1 Sample Collection and Coding 3.3.2 Anamnesis and Identity Record 3.3.3 Sample Collection 3.3.4 Direct Microscopic Observation 3.3.5 Fungal Culture 3.3.6 Macroscopic Observation of SDA Agar Plates 3.3.7 Microscopic Observation of SDA Agar Plates 3.4 Data Analysis IV RESULTS 4.1 Cats 4.2 Direct Examination 4.3 Identification of Isolated Dermatophytes 4.4 Prevalence of Dermatophytes IV DISCUSSION 5.1 Lesions in Sampled Cats 5.2 Diagnostic Approaches 5.3 Prevalence of Dermatophytosis in Cats 5.3.1 Conditions Influencing Prevalence 5.3.2 Reasons for the Discrepancies 5.4 Microsporum canis and Trichophyton verrucosum	II	2.1 2.2 2.3	Cats Diseases Caused by Fungi in Cat Diagnosis of Fungal Infection in Cat 2.3.1 Wood Lamp Examination 2.3.2 Direct Microscopic Examination 2.3.3 Fungal Culture	
3.1 Time and Place 3.2 Study Animals and Materials 3.3 Methodology 3.3.1 Sample Collection and Coding 3.3.2 Anamnesis and Identity Record 3.3.3 Sample Collection 3.3.4 Direct Microscopic Observation 3.3.5 Fungal Culture 3.3.6 Macroscopic Observation of SDA Agar Plates 3.3.7 Microscopic Observation of SDA Agar Plates 3.4 Data Analysis IV RESULTS 4.1 Cats 4.2 Direct Examination 4.3 Identification of Isolated Dermatophytes 4.4 Prevalence of Dermatophytes 4.4 Prevalence of Dermatophytes 5.1 Lesions in Sampled Cats 5.2 Diagnostic Approaches 5.3 Prevalence of Dermatophytosis in Cats 5.3.1 Conditions Influencing Prevalence 5.3.2 Reasons for the Discrepancies 5.4 Microsporum canis and Trichophyton verrucosum			•	
4.1Cats104.2Direct Examination134.3Identification of Isolated Dermatophytes144.4Prevalence of Dermatophytes20IV DISCUSSION225.1Lesions in Sampled Cats225.2Diagnostic Approaches235.3Prevalence of Dermatophytosis in Cats245.3.1Conditions Influencing Prevalence255.3.2Reasons for the Discrepancies255.4Microsporum canis and Trichophyton verrucosum26	111	3.1 3.2 3.3	Time and Place Study Animals and Materials Methodology 3.3.1 Sample Collection and Coding 3.3.2 Anamnesis and Identity Record 3.3.3 Sample Collection 3.3.4 Direct Microscopic Observation 3.3.5 Fungal Culture 3.3.6 Macroscopic Observation of SDA Agar Plates 3.3.7 Microscopic Observation of SDA Agar Plates	6 6 6 6 7 8 8
 5.1 Lesions in Sampled Cats 5.2 Diagnostic Approaches 5.3 Prevalence of Dermatophytosis in Cats 5.3.1 Conditions Influencing Prevalence 5.3.2 Reasons for the Discrepancies 5.4 Microsporum canis and Trichophyton verrucosum 	IV	4.1 4.2 4.3	Cats Direct Examination Identification of Isolated Dermatophytes	10 13 14
	IV	5.15.25.3	Lesions in Sampled Cats Diagnostic Approaches Prevalence of Dermatophytosis in Cats 5.3.1 Conditions Influencing Prevalence 5.3.2 Reasons for the Discrepancies Microsporum canis and Trichophyton verrucosum	22 23 24 25 25 26





10		Hak 1. Di
0.0	0.0	_ 0
0.0		2.8
0 0	P	01 0
- CO	TO	cCipta Dilindungi Undang-undan Dilarang mengutip sebagian atau
T (F)		3 5
32 E		m m
3 5.	S.	9 2
2 2	P	3 =
3 4		7 6
00	_	00 =
E 2	83	5.3
3 0	3	7-00
EX	× 2	(e =
3 3	100	0 5
S 6	=	OT D
5 5	-	01 D
0.00	드	27 mg
0 =	_	7 7
	6	
5 5	TO	# 5
70 00	LD.	0 0
3 0	=	
70 0		CD OLD
6 7	m	=
2 0	0	=
38	-	=
b. Pengutipan tidak merugikan kepentingan yang wajar IPB University. . Dilarang mengumumkan dan memperbanyak sebagian atau seluruh karya tulis ini dalam bentuk apapun tanpa izin IPB University.	Pengutipan hanya untuk kepentingan pendidikan, penelitian, penulisan karya ilmiah, penyusunan laporan, penulisan kritik atau	lak Cipta Dilindungi Undang-undang . Dilarang mengutip sebagian atau seluruh karya tulis ini tanpa mencantumkan dan menyebutkan sumber
× 5	3	6
20 V	0	7
V1 (1)		61
0 00	=	- LD
0.00	1	C
m 3	3	
20	-	01
3 9		=
		_
7	D	SUF
20 00	=	
0 Z	3	
2 3	-	3
5 9		Œ
= 0	3	5
2 3		Si .
6	=	=
7	8	8
01	3	=
-	~	
=	00	S)
	3	-
5.	D	Q.
3		=
-	3	_
01		3
		CD
3	-	3
5		(T)
00-	~	5
- 2	E	8
=		
0	3	-
70		
a)	2	=
TO		2
	D	rb
-	0	77
0.5	0.5	
3	3	
D	73	
00	ĕ	
	3	
3		
	10	
(U)	3	
_	8	
2	=	
5	=	
10	03	
US.	-	
2	B	
	-	

r		4
F		н
	U	ч
Н		9
t	Ţ	IJ
1	ò	
r		4
P	7	
Е	٥	
٤	٥,	
P	4	
C	D	
Е	ą	
	×	
Ľ	Z,	
k	=	
4	J	

10	5.6 C	Challenges in Controlling Dermatophytosis in Shelters	27
	5.7 A	wareness of Zoonotic Risks in Shelter	27
	5.8 I	Limitation of the Study	29
VI	CONCLU	JSIONS AND SUGGESTIONS	30
9	6.1 C	Conclusion	30
Hak	6.2 S	uggestions	30
RE	FERENCE	E	31
AP	PENDIX		37
	OGRAPHY	7	44
7 Ba			
niv			
ersi			
4			

n sumber : unan laporan, penulisan kritik atau tinjauan suatu masalah



LIST OF TABLES

Clinical information of cats for dermatophytes examination

9

2	Lesions scoring of the sampled cats	11
3	Results of Wood's Lamp test, direct microscopic examination, trichogram, and fungal culture	12
	LIST OF FIGURES	
1	British Shorthair (A); Persian (B)	3
2	Skin changes seen in cats infected by dermatophytosis	4
3	Green fluorescent light appears under a Wood's lamp examination	4
4	Microscopic examination of hairs (10x10). Infected hairs are paler and	
	border than normal hairs	5
5	Diagram of the brief procedure of dermatophytes identification in shelter	
	cats	8
6	Wood's lamp examination results on cats	15
7	Direct examination of samples from skin scrap and hair pluck at 40×10	
_	magnification	15
8	Macroscopic observation of <i>Trichophyton verruscum</i> from C08B on SDA	1.0
0	M:	16
9	Microscopic image of <i>Trichophyton verrucosum</i> (magnification 40x10) from C08B on SDA	17
10	Macroscopic observation of <i>Trichophyton verrucosum</i> from C08B on	1 /
10	PDA	17
11	Microscopic image of <i>Trichophyton verrucosum</i> (magnification	1 /
11	40×10) from C08B on PDA	18
12	Macroscopic observation of <i>Microsporum canis</i> from C18A on SDA	19
13	Macroscopic observation of <i>Microsporum canis</i> from C18B on SDA	19
14	Macroscopic observation of <i>Microsporum canis</i> from C18B2 on SDA	20
15	Microscopic observation of <i>Microsporum canis</i> (magnification 40×10)	
	from C18A, C18B, and C18B2 on SDA	20
16	Macroscopic observation of Microsporum canis colonies from C18B2 on	
	PDA	21
17	Microscopic observation of Microsporum canis (40×10 magnification)	
	from C18B2 from PDA	22

1



