

ISBN: 978-623-90888-0-4



# Proceedings

## Internasional Seminar on Tropical Horticulture 2018: Horticulture for The Quality of Life



December 10, 2018  
Bogor, Indonesia  
IPB International Convention Center

Published by:  
PKHT (Center for Tropical Horticulture Studies), IPB University  
Bogor, April 2019



# **PROCEEDINGS**

## **International Seminar on Tropical Horticulture 2018**

### ***Horticulture for The Quality of Life***

December 10<sup>th</sup>, 2018  
IPB International Convention Center- IPB ICC  
Bogor, Indonesia

Published by:  
**Pusat Kajian Hortikultura Tropika LPPM – IPB**  
(Center for Tropical Horticulture Studies)  
**2019**

**Title:**

Proceedings The International Seminar on Tropical Horticulture 2018

**Theme:**

Horticulture for The Quality of Life

**Organizing Committee:**

Chairperson	: Awang Maharijaya
Vice Chairperson	: Kusuma Darma
Secretary	: Netty Tinaprila
Treasurer	: Heri Harti, Member
Event	: Andi Azhari Putra
	: Ayuni Nuramalina
	: Maxymiland Leiwakabesy
	: Pritha K.H
Public Relation	: Galuh Hanifatiha
	: Ita Aprilia
IT and Website	: Ferdhi Isnani Nuryana
Secretariat	: Rika Lesmawati
	: Rena Destriani
	: Dedeh Saptri

**Sterring Committee:**

1. Syafrida Manuwoto
2. Sobir
3. Darda Effendi
4. Muhammad Rahmad Suhartanto
5. Muhammad Syukur
6. Y. Aris Purwanto

**Reviewer:**

1. Kusuma Darma
2. Heri Harti
3. Endang Gunawan
4. Naekman Naibaho

**Editor:**

Awang Maharijaya

**Published by:**

Pusat Kajian Hortikultura Tropika (PKHT) - LPPM, IPB

Center for Tropical Horticulture Studies

Kampus IPB Baranangsiang, Jalan Pajajaran Tegallega, Bogor Tengah,  
Bogor, 16129, West Java, Indonesia

Telp/Fax: (0251) 8326881

email: [pkht.ipb@gmail.com](mailto:pkht.ipb@gmail.com)

Publish online in PKHT's website <http://pkht.ipb.ac.id/>

**ISBN** 978-623-90888-0-4

© 2018, ALL RIGHT RESERVED No part of this publication may be reproduced, store in a retrieval system, or by any means, electronic, mechanical or photocopying without the prior formal permission of the authors.

# FOREWORD

International Seminar on Tropical Horticulture 2018: Horticulture for The Quality of Life was held in IPB International Convention Center, Bogor, Indonesia, December 10, 2018. This seminar organized by Center for Tropical Horticulture Studies (PKHT), IPB University and supported by Center of Excellence in University (PUI-PT), Ministry of Research, Technology and Higher Education.

We're very glad to know the fact that the seminar displayed a very wide discussion about tropical horticulture with delegates from 5 countries (Japan, Taiwan, Thailand, China, and Indonesia) as keynote speech and participants. 31 papers were selected to be included in this proceeding from 23 oral presentation and 38 poster presentation.

This proceeding is contained of three sub chapter, that is ornamental plants, fruits, and vegetables. There are 5 papers of ornamental plants, 8 papers of fruits chapter and 17 papers of vegetables chapter

We wish to thank Prof. Slamet Susanto, Prof. Sanesuki Kawabata, Prof. Masayoshi Shigyo, Prof. Sobit, Prof. Ming-Tsair Cahn, Mr. Rahmansyah Darmawan being keynote speech at this international seminar and all participants for very lively atmosphere during and after the seminar.

Bogor, July 2019

Editor

Dr. Awang Maharijaya

## **Table of Contents**

<b>Foreword</b>	<b>iii</b>
<b>Table of Contents</b>	<b>iv</b>
<b>List of Paper</b>	<b>v</b>
<b>Schedule of The Seminar</b>	<b>viii</b>
<b>The Committee</b>	<b>ix</b>
<b>List of Oral Presentation Titles</b>	<b>x</b>
<b>List of Poster Presentation Titles</b>	<b>xii</b>
<b>List of Participants</b>	<b>xiii</b>

## LIST OF PAPERS

### Ornamental Plants

<b>Orchid and Pitcher Plant of Batam Botanic Gardens Collections</b> Yupi Isnaini and Hartutiningsih M-Siregar .....	3
<b>Developing Descriptor List for Hoya Plant</b> Sri Rahayu.....	13
<b>Biomass Production and Quality of Orange Jessamine (<i>Murraya paniculata</i> [L.] Jack) Leaves at Two Harvest Interval</b> Noorwitri Utami, Sandra Arifin Aziz and Maya Melati.....	26
<b>Effect of Coating Compounds and Altitude of Storage Location on Satoimo Corm Quality (<i>Colocasia esculenta</i> (L) Scott var <i>Antiquorum</i>)</b> Winda Nawfetrias, Delvi Maretta, Dwi Pangesti H and Hendra Asmara .....	37
<b>The Influence of Scarification and Growing Media on Germination of Palm Squirrel's Tailed Seed (<i>Wodyetia bifurcata</i>)</b> Chitra Priatna, Ronald Bunga Mayang and Djoko Mulyono .....	43

### Fruits

<b>Effect of Paclobutrazol Concentration and Wire Winding on off Season Flowering Induction in Orange Plants</b> Isna Tustiyani, Asi Yasinta and Siti Syarah Maesyaroh .....	51
<b>The Effectiveness on Different Kind of Growth Regulators at The Stage of Seed Multiplication Technique with Turn Off The Growing Point of the Tanduk Banana Corm (<i>Musa paradisiaca formatypica</i>)</b> Eltis Panca Ningsih and Imas Rohmawati .....	56
<b>Identification Of Bioactive Compounds And Insecticidal Potential of JC Citrus Peel Extract To Citrus Psyllid <i>Diaphorina citri</i> and Citrus Aphids <i>Aphis gossypii</i></b> S. Wuryantini, Harwanto and R. A. Yudhistira .....	62
<b><i>Trichoderma</i> sp. as Diplodia Disease Control for Citrus in Swamp Area</b> Susi Wuryantini, M. Taufik, Dina Agustina and Mutia Erti .....	71
<b>Effects of Botanical Pesticides On The Growth Of <i>Sclerotinia sclerotiorum</i></b> Dina Agustina, Sriwidyarningsih, and Unun Triasih .....	76
<b>Biological Control of Post harvest disease of Mango by using antagonistic</b> Roza Yunita, Roedhy Poerwanto and Suryo Wiyono .....	82
<b>Potential of Processing Technology Astano Pummelo Citrus Peel in Jambi Province</b> Dewi Novalinda and Desi Hernita .....	89



**Optimization of Papaya Callina Soft Candy Formulation (*Carica Papaya* L.) Using The Response Surface Method**

Hisworo Ramdani, Yoni Atma, Maulina Nuroktaviani Rahayu ..... 94

**Vegatables**

**Characterization of Morphology and Potencial of Garlic Genetic Resources in Jambi Province**

Desi Hernita, Eva Salvia and Jon Hendri ..... 108

**Characterization of 12 Introduced Garlic (*Allium sativum* L.) Genotypes on Indonesia Vegetable Research Institute, Lembang, West Java**

Eti Heni Krestini, Catur Hermanto, Nazly Aswani ..... 115

**Effect of Varieties and Use of Mulch on Garlic Bulb Quality**

Suwarni Tri Rahayu and Levianny PS ..... 122

**Performance of Some Genotypes of Shallot in Dry Acid Soil, South Lampung**

Rismawita Sinaga, Nurmawati Waluyo, Iteu M. Hidayat and Gohan Octora Manurung... 129

**Effectiveness of Decontamination Treatments for Tissue Culture Initiation of Shallots (*Allium ascalonicum* L.) and Garlic (*Allium sativum*) Explants**

Asih K. Karjadi and Neni Gunaeni ..... 135

**The effect of antiviral Ribavirin on Proliferation of Explants Shallots cv. Trisula and cv. Bima Brebes cv. Trisula and cv. Bima Brebes**

Asih K. Karjadi and Neni Gunaeni ..... 142

**Genetic Variability and Agronomic Characters of Bambara Groundnut (*Vigna Subterranea* L. Verdc.) Lines Derived from Pure Line Selection of Sukabumi Landrace**

Yuliawati, Yudiwanti Wahyu EK, Memen Surahman and Arifah Rahayu ..... 152

**Move to Organic Farming Systems: Chemical and Physical Soil Properties of Organic and Conventional Vegetables Reasons**

Sukristiyonubowo, Damasus Riyanto and Sugeng Widodo ..... 162

**Potato Breeding Technology in Kayu Aro, Jambi**

Lutfi Izhar, Lindayanti, Syafriedi and Salwati ..... 170

**Potato Crop Simulation Model**

Salwati and Lutfi Izhar ..... 187

**Production of Potato Seed on Many Nitrogen Rates and Its Changed on Comparison NO<sub>3</sub> and NH<sub>4</sub> of AB Mix Fertilizer**

Meksy Dianawati ..... 184

**Construction and Evaluation of DNA-A and DNA-B PYLCV (Pepper Yellow Leaf Curl Virus) In *Agrobacterium* Plasmid**

Ula Aulia Fitriani, Tati Kristianti and Sony Suhandono ..... 190

<b>Study of The Use of Varieties and Fertilizer on The Growth and Development of Virus Disease in Hot Chili (<i>Capsicum frutescens</i>)</b> Eli Korlina, Riza Ulil Fitria and Sri Zunaini Saadah .....	197
<b>Management and Utilization of Biological Resources Control of Important Plant Disturbing Organisms on Chili Pepper</b> Neni Gunaeni and Astri W. Wulandari .....	212
<b>Correlation between Virus Attacks at the Several Phases of Growth with the Yield of Chili (<i>Capsicum Annuum</i> L.) in Lowland</b> Elly Kesumawati, Sabaruddin, Muhammad Asdhani and Sofyan .....	221
<b>The Technology of Red Chili Processing to Improve The Quality of Chili in Jambi Province</b> Linda Yanti, Dewi Novalinda and Desi Hernita .....	232
<b>Identification of Amino Acid, fatty acid, mineral content and respiration rate of straw mushroom in different stage of maturity</b> Resa Setia Adiandri, Nikmatul Hidayah and Sigit Nugraha .....	239
<b>Impact Increasing Input and Output Prices Toward Competitiveness of Chili in Banten Province</b> Viktor Siagian .....	249

# **PROCEEDINGS**

## **International Seminar on Tropical Horticulture 2018**

### ***Horticulture for The Quality of Life***

December 10<sup>th</sup>, 2018  
IPB International Convention Center- IPB ICC  
Bogor, Indonesia

Published by:  
**Pusat Kajian Hortikultura Tropika LPPM – IPB**  
(Center for Tropical Horticulture Studies)  
**2019**

**Title:**

Proceedings The International Seminar on Tropical Horticulture 2018

**Theme:**

Horticulture for The Quality of Life

**Organizing Committee:**

Chairperson	: Awang Maharijaya
Vice Chairperson	: Kusuma Darma
Secretary	: Netty Tinaprila
Treasurer	: Heri Harti, Member
Event	: Andi Azhari Putra
	: Ayuni Nuramalina
	: Maxymiland Leiwakabesy
	: Pritha K.H
Public Relation	: Galuh Hanifatiha
	: Ita Aprilia
IT and Website	: Ferdhi Isnani Nuryana
Secretariat	: Rika Lesmawati
	: Rena Destriani
	: Dedeh Saptri

**Sterring Committee:**

1. Syafrida Manuwoto
2. Sobir
3. Darda Effendi
4. Muhammad Rahmad Suhartanto
5. Muhammad Syukur
6. Y. Aris Purwanto

**Reviewer:**

1. Kusuma Darma
2. Heri Harti
3. Endang Gunawan
4. Naekman Naibaho

**Editor:**

Awang Maharijaya

**Published by:**

Pusat Kajian Hortikultura Tropika (PKHT) - LPPM, IPB

Center for Tropical Horticulture Studies

Kampus IPB Baranangsiang, Jalan Pajajaran Tegallega, Bogor Tengah,  
Bogor, 16129, West Java, Indonesia

Telp/Fax: (0251) 8326881

email: [pkht.ipb@gmail.com](mailto:pkht.ipb@gmail.com)

Publish online in PKHT's website <http://pkht.ipb.ac.id/>

**ISBN** 978-623-90888-0-4

© 2018, ALL RIGHT RESERVED No part of this publication may be reproduced, store in a retrieval system, or by any means, electronic, mechanical or photocopying without the prior formal permission of the authors.

# FOREWORD

International Seminar on Tropical Horticulture 2018: Horticulture for The Quality of Life was held in IPB International Convention Center, Bogor, Indonesia, December 10, 2018. This seminar organized by Center for Tropical Horticulture Studies (PKHT), IPB University and supported by Center of Excellence in University (PUI-PT), Ministry of Research, Technology and Higher Education.

We're very glad to know the fact that the seminar displayed a very wide discussion about tropical horticulture with delegates from 5 countries (Japan, Taiwan, Thailand, China, and Indonesia) as keynote speech and participants. 31 papers were selected to be included in this proceeding from 23 oral presentation and 38 poster presentation.

This proceeding is contained of three sub chapter, that is ornamental plants, fruits, and vegetables. There are 5 papers of ornamental plants, 8 papers of fruits chapter and 17 papers of vegetables chapter

We wish to thank Prof. Slamet Susanto, Prof. Sanesuki Kawabata, Prof. Masayoshi Shigyo, Prof. Sobit, Prof. Ming-Tsair Cahn, Mr. Rahmansyah Darmawan being keynote speech at this international seminar and all participants for very lively atmosphere during and after the seminar.

Bogor, July 2019

Editor

Dr. Awang Maharijaya

## **Table of Contents**

<b>Foreword</b>	<b>iii</b>
<b>Table of Contents</b>	<b>iv</b>
<b>List of Paper</b>	<b>v</b>
<b>Schedule of The Seminar</b>	<b>viii</b>
<b>The Committee</b>	<b>ix</b>
<b>List of Oral Presentation Titles</b>	<b>x</b>
<b>List of Poster Presentation Titles</b>	<b>xii</b>
<b>List of Participants</b>	<b>xiii</b>

## LIST OF PAPERS

### Ornamental Plants

<b>Orchid and Pitcher Plant of Batam Botanic Gardens Collections</b> Yupi Isnaini and Hartutiningsih M-Siregar .....	3
<b>Developing Descriptor List for Hoya Plant</b> Sri Rahayu.....	13
<b>Biomass Production and Quality of Orange Jessamine (<i>Murraya paniculata</i> [L.] Jack) Leaves at Two Harvest Interval</b> Noorwitri Utami, Sandra Arifin Aziz and Maya Melati.....	26
<b>Effect of Coating Compounds and Altitude of Storage Location on Satoimo Corm Quality (<i>Colocasia esculenta</i> (L) Scott var Antiquorum)</b> Winda Nawfetrias, Delvi Maretta, Dwi Pangesti H and Hendra Asmara .....	37
<b>The Influence of Scarification and Growing Media on Germination of Palm Squirrel's Tailed Seed (<i>Wodyetia bifurcata</i>)</b> Chitra Priatna, Ronald Bunga Mayang and Djoko Mulyono .....	43

### Fruits

<b>Effect of Paclobutrazol Concentration and Wire Winding on off Season Flowering Induction in Orange Plants</b> Isna Tustiyani, Asi Yasinta and Siti Syarah Maesyaroh .....	51
<b>The Effectiveness on Different Kind of Growth Regulators at The Stage of Seed Multiplication Technique with Turn Off The Growing Point of the Tanduk Banana Corm (<i>Musa paradisiaca formatypica</i>)</b> Eltis Panca Ningsih and Imas Rohmawati .....	56
<b>Identification Of Bioactive Compounds And Insecticidal Potential of JC Citrus Peel Extract To Citrus Psyllid Diaphorina Citri and Citrus Aphids Aphis Gossypii</b> S. Wuryantini, Harwanto and R. A. Yudhistira .....	62
<b><i>Trichoderma</i> sp. as Diplodia Disease Control for Citrus in Swamp Area</b> Susi Wuryantini, M. Taufik, Dina Agustina and Mutia Erti .....	71
<b>Effects of Botanical Pesticides On The Growth Of <i>Sclerotinia Sclerotiorum</i></b> Dina Agustina, Sriwidyarningsih, and Unun Triasih .....	76
<b>Biological Control of Post harvest disease of Mango by using antagonistic</b> Roza Yunita, Roedhy Poerwanto and Suryo Wiyono .....	82
<b>Potential of Processing Technology Astano Pummelo Citrus Peel in Jambi Province</b> Dewi Novalinda and Desi Hernita .....	89



**Optimization of Papaya Callina Soft Candy Formulation (*Carica Papaya* L.) Using The Response Surface Method**

Hisworo Ramdani, Yoni Atma, Maulina Nuroktaviani Rahayu ..... 94

**Vegatables**

**Characterization of Morphology and Potencial of Garlic Genetic Resources in Jambi Province**

Desi Hernita, Eva Salvia and Jon Hendri ..... 108

**Characterization of 12 Introduced Garlic (*Allium sativum* L.) Genotypes on Indonesia Vegetable Research Institute, Lembang, West Java**

Eti Heni Krestini, Catur Hermanto, Nazly Aswani ..... 115

**Effect of Varieties and Use of Mulch on Garlic Bulb Quality**

Suwarni Tri Rahayu and Levianny PS ..... 122

**Performance of Some Genotypes of Shallot in Dry Acid Soil, South Lampung**

Rismawita Sinaga, Nurmawati Waluyo, Iteu M. Hidayat and Gohan Octora Manurung... 129

**Effectiveness of Decontamination Treatments for Tissue Culture Initiation of Shallots (*Allium ascalonicum* L.) and Garlic (*Allium sativum*) Explants**

Asih K. Karjadi and Neni Gunaeni ..... 135

**The effect of antiviral Ribavirin on Proliferation of Explants Shallots cv. Trisula and cv. Bima Brebes cv. Trisula and cv. Bima Brebes**

Asih K. Karjadi and Neni Gunaeni ..... 142

**Genetic Variability and Agronomic Characters of Bambara Groundnut (*Vigna Subterranea* L. Verdc.) Lines Derived from Pure Line Selection of Sukabumi Landrace**

Yuliawati, Yudiwanti Wahyu EK, Memen Surahman and Arifah Rahayu ..... 152

**Move to Organic Farming Systems: Chemical and Physical Soil Properties of Organic and Conventional Vegetables Reasons**

Sukristiyonubowo, Damasus Riyanto and Sugeng Widodo ..... 162

**Potato Breeding Technology in Kayu Aro, Jambi**

Lutfi Izhar, Lindayanti, Syafriedi and Salwati ..... 170

**Potato Crop Simulation Model**

Salwati and Lutfi Izhar ..... 187

**Production of Potato Seed on Many Nitrogen Rates and Its Changed on Comparison NO<sub>3</sub> and NH<sub>4</sub> of AB Mix Fertilizer**

Meksy Dianawati ..... 184

**Construction and Evaluation of DNA-A and DNA-B PYLCV (Pepper Yellow Leaf Curl Virus) In *Agrobacterium* Plasmid**

Ula Aulia Fitriani, Tati Kristianti and Sony Suhandono ..... 190

<b>Study of The Use of Varieties and Fertilizer on The Growth and Development of Virus Disease in Hot Chili (<i>Capsicum frutescens</i>)</b> Eli Korlina, Riza Ulil Fitria and Sri Zunaini Saadah .....	197
<b>Management and Utilization of Biological Resources Control of Important Plant Disturbing Organisms on Chili Pepper</b> Neni Gunaeni and Astri W. Wulandari .....	212
<b>Correlation between Virus Attacks at the Several Phases of Growth with the Yield of Chili (<i>Capsicum Annuum</i> L.) in Lowland</b> Elly Kesumawati, Sabaruddin, Muhammad Asdhani and Sofyan .....	221
<b>The Technology of Red Chili Processing to Improve The Quality of Chili in Jambi Province</b> Linda Yanti, Dewi Novalinda and Desi Hernita .....	232
<b>Identification of Amino Acid, fatty acid, mineral content and respiration rate of straw mushroom in different stage of maturity</b> Resa Setia Adiandri, Nikmatul Hidayah and Sigit Nugraha .....	239
<b>Impact Increasing Input and Output Prices Toward Competitiveness of Chili in Banten Province</b> Viktor Siagian .....	249

# Biomass Production and Quality of Orange Jessamine (*Murraya paniculata* [L.] Jack) Leaves at Two Harvest Interval

Noorwitri Utami<sup>1\*</sup>, Sandra Arifin Aziz<sup>2</sup> and Maya Melati<sup>2</sup>

<sup>1</sup> Center for Agricultural Production Technology, Agency for the Assessment and Application of Technology, Serpong 15314, Indonesia

<sup>2</sup> Departement of Agronomy and Horticulture, Faculty of Agriculture, Bogor Agricultural University, Jalan Meranti, Bogor 16680, Indonesia

\*Corresponding author: noorwitri.utami@bppt.go.id

## Abstract

Orange jessamine is a medicinal plant that has potential as an antioxidant and until now information on the cultivation and quality of its leaves active compound still limited. This research was conducted at Bogor Agricultural University experimental station (Indonesia), from April to October 2013. The objectives of this research was to study the effect of harvest interval on biomass production and bioactive content. The experiment was laid out in a completely randomized block design with single factor, i.e. 5 and 12 weeks harvest interval, with three replications. Data were analyzed using t student test. Plants harvested every 12-weeks gives best yield and regrowth for orange jessamine. Protein content and PAL activity were increased in the next harvest, either at 5 or 12-weeks harvest interval. Plants harvested every 5-weeks for three harvests had no significantly different on phenol and flavonoid production with 12-weeks harvest interval for one harvest.

---

Keywords: flavonoid, harvest interval, *Murraya paniculata*, phenolic, phenylalanine ammonia-lyase (PAL)

## 1. Introduction

Orange jessamine (*Murraya paniculata* [L.] Jack) is a medicinal plant of the family Rutaceae, in effective oral doses is safe to use (Gautam *et al.*, 2012a) and has the potential to reduce blood cholesterol levels (Pane, 2010), antiobesity (Iswantini *et al.*, 2011), antidiabetic (Gautam *et al.*, 2012b), antifertility (Xiao dan Wang, 1991), antidiarrheal (Rahman *et al.*, 2010), anti-inflammatory and *antinociceptive* (Wu *et al.*, 2010; Podder *et al.*, 2011) and antioxidant (Gautam *et al.*, 2012c; Paramaguru *et al.*, 2012).

A plant has the potential the potential as an antioxidant characterized by the presence of polyphenols (Lugasi *et al.*, 2003). Polyphenols are divided into two main groups namely phenolic and flavonoids, which create 1/3 to 2/3 of all antioxidants (Tapiero *et al.*, 2002). The antioxidant effect of orange jessamine leaves might be due to the phenolic content of it leaves. Orange jessamine leaves contain steroids, saponins, flavonoids, tannins and alkaloids (Syahadat and Aziz, 2012). Flavonoids are part of poliphenols which are known to have properties as free radical scavenging, hydrolysis and oxidative enzyme inhibitors and work as anti-inflammatory (Pourmorad *et al.*, 2006).

All phenolic compounds are produced from the phenylpropanoid pathway. Phenylalanine ammonia lyase (PAL) is an important enzyme in the production of phenolic compounds. The level of PAL activity depends on the genotype, age and development stage, organs, as well as plant tissue types (Camm and Towers, 1973). In addition, PAL activity is also influenced by various factors including light, temperature, growth regulators, RNA inhibitors, protein synthesis, wounding and nutrients (Jones, 1984).

Flavonoids are derivative products of the phenylpropanoid pathway. An important step in flavonoid biosynthesis is the condensation of malonyl CoA molecules with one molecule p-coumaroyl-CoA to C15 chalcone intermediate (naringenin chalcone), this reaction is catalyzed by chalcone synthase and subsequently transformed into various flavonoid products (Cheng *et al.*, 2009). Anthocyanins are part of flavonoid compounds and have antioxidant effects to protect the heart (cardioprotective) (Ververidis *et al.*, 2007).

Orange jessamine has been widely used, but information about crop cultivation is still limited, particularly harvest interval as it will determine the active ingredient in it. Proper harvest settings will be useful to get optimal results with maximum quality. The objective of this research was to study the effect of harvest intervals on biomass production and its bioactive content.

## 2. Materials and Method

The research was conducted from April to October 2013 in the Cikarawang IPB experimental garden, Dramaga, Bogor. The materials and tools used in this study include 18-month old orange jessamine, agricultural lime (85% CaCO<sub>3</sub>), laying hen manure, rock phosphate, husk ash, Heraeus Labofuge-400R centrifuge, freeze dryer Flexy-Dry™ MP, Waterbath Eyla SB-24, and Shimadzu UV-1800 spectrophotometer.

The study was conducted using a randomized block design of one treatment factor, namely the harvest interval. The harvest interval consists of two levels, namely 5 and 12 weeks. Each treatment consists of six plants. The experiment was carried out using plots with a size of 2 m x 1 m and a distance between plot 50 cm. Basic treatment in the form of agricultural lime and laying hen manure is given before planting seedlings in the field. Agricultural lime (2 ton/ha) is given by way of evenly distributed on land 4 weeks before planting (WBP). Laying hen manure (5 kg/planting hole) is given 1 WBP. Rock phosphate (0.45 kg) and rice-hull ash (2 kg) are given around the planting hole together at planting. Planting is done by transferring seedling from polybags to plots. The spacing used is 1 m with a depth of 30 cm. Flowers and fruits from generative phase were removed from seedlings.

Uniformity of plant height was carried out four weeks after planting at a height of 75 cm from the ground. Harvesting is done by cutting the plants at a cutting height of 75 cm. Harvesting at 5-week intervals is carried out at 9, 14 and 19 weeks after planting (WAP); while harvesting at 12-week intervals was carried out at 16 and 28 WAP (Table 1).

*Table 1. Harvest schedule of Orange Jessamine during the observation*

Harvest intervals (weeks)	4	9	14	16	19	28
	.....WAP (weeks after planting).....					
5	√*	√	√		√	
12	√*			√		√

Note: √ = harvest, \* = harvest is not included in data processing.

The variables observed included biomass production, leaves quality, and secondary metabolite production. The biomass observed was leaves fresh weight, stem-branch fresh weight, total fresh weight (leaves weight + stem-branch weight), leaves dry weight, stem-branch dry weight, total dry weight (leaves dry weight + stem-branch dry). Leaves quality variables were observed in medium leaves (leaves after the fifth leaf from shoots), including: protein content (Lowry *et al.*, 1951), PAL (Dangcham *et al.*, 2008), phenolic content which calculated gallic acid equivalents (Waterhouse, 2002), flavonoid levels which calculated the quercetin equivalent (Chang *et al.*, 2002). The production of secondary metabolites observed included phenolic production and flavonoid production. The production of secondary metabolites is calculated as a result of multiplication of leaves biomass with its levels. Changes in biomass, leaf quality and secondary metabolite production were observed at 9, 14 and 19 WAP at the 5-week harvest interval; and at 16 and 28 WAP at the harvest interval every 12-weeks.

The measurement data were analyzed using student t-test to determine the harvest interval which gave the best response to the observed variables.

### 3. Results and Discussion

Biomass production and bioactive content of orange jessamine are influenced by cultivation practices, including harvest intervals. Harvest intervals affect biomass production, nutritional value, and the ability to regrow plants afterward (Man and H Wiktorsson, 2003).

#### Effect of harvest intervals on biomass production (g)

The second harvest at both the five-week and 12-week harvest intervals gave a very significant increase in leaves fresh weight, stem fresh weight and total fresh weight compared to the first harvest (Figures 1a, 2a, and 3a), this was due to loss of apical dominance which stimulated branch formation, more lateral branches so that biomass production rises in the second harvest. However, in the third harvest at the five-week harvest interval, poor crop regrowth was characterized by decreased biomass of orange jessamine (Figures 1a, 2a, and 3a). Orange jessamine harvesting at the five-week harvest interval in three harvests gave a cumulative result that was not significantly different from the first harvest of the 12-week harvest interval on biomass production components including leaves fresh weight, stem fresh weight and total fresh weight (Figures 1a, 2a and 3a). Short harvest intervals cause the opportunity for plants to accumulate photosynthates to be shorter, while at longer leaves harvest intervals plants can grow longer to accumulate photosynthetic results so that plants can grow optimally.

This result was same with the previous experiment that the highest fresh and dry forage production of torbangun (*Coleus amboinicus*) plants was obtained at 60-day cutting intervals, followed by 40, 30-day cutting intervals and the lowest at 50-day cutting intervals (Sajimin *et al.*, 2011).

The dry weight of plants reflects the accumulation of organic compounds that have been synthesized by plants from inorganic compounds, especially water and carbon-dioxide. The dry weight gain of plants is also a contribution of nutrients that have been absorbed by the roots. The dry weight of plants is a result of the efficiency of absorption and utilization of solar radiation available throughout the cropping period by plant canopy (Kastono *et al.*, 2005). The dry weight of leaves, stem and branch dry weight, also total dry weight in the second harvest at the five-week and 12-week harvest intervals increased significantly compared to the first harvest, while leaves dry weight, stem-branch dry weight and total dry weight of orange jessamine in the third harvest at the five-week harvest interval decreased compared with results in the second harvest and not significantly different from dry weight in the first harvest (Figures 1b, 2b, and 3b).

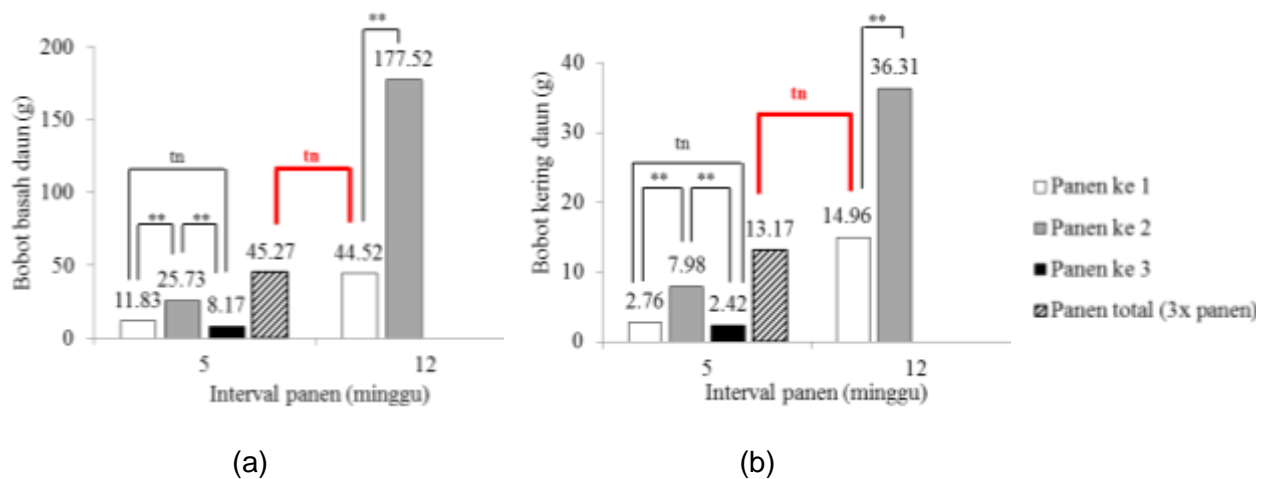


Figure 1. Fresh (a) and dry leaves weight (b) of orange jessamine at 5 and 12-weeks harvest interval; \*\*: significantly different at  $P \leq 0.01$ , ns: not significant at  $P > 0.05$ .

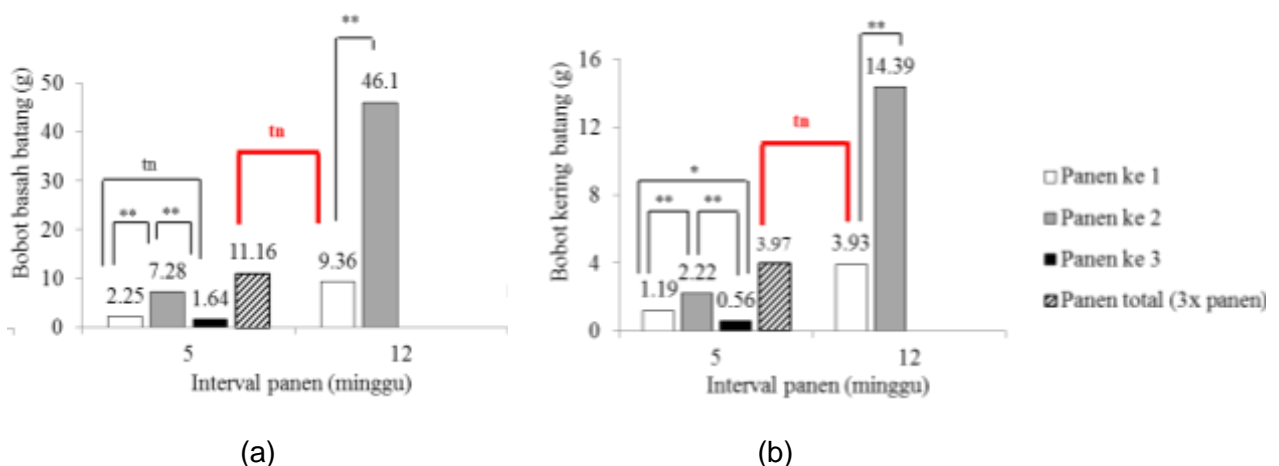


Figure 2. Fresh (a) and dry stem weight (b) of orange jessamine at 5 and 12-weeks harvest interval; \*\*: significantly different at  $P \leq 0.01$ , ns: not significant at  $P > 0.05$ .

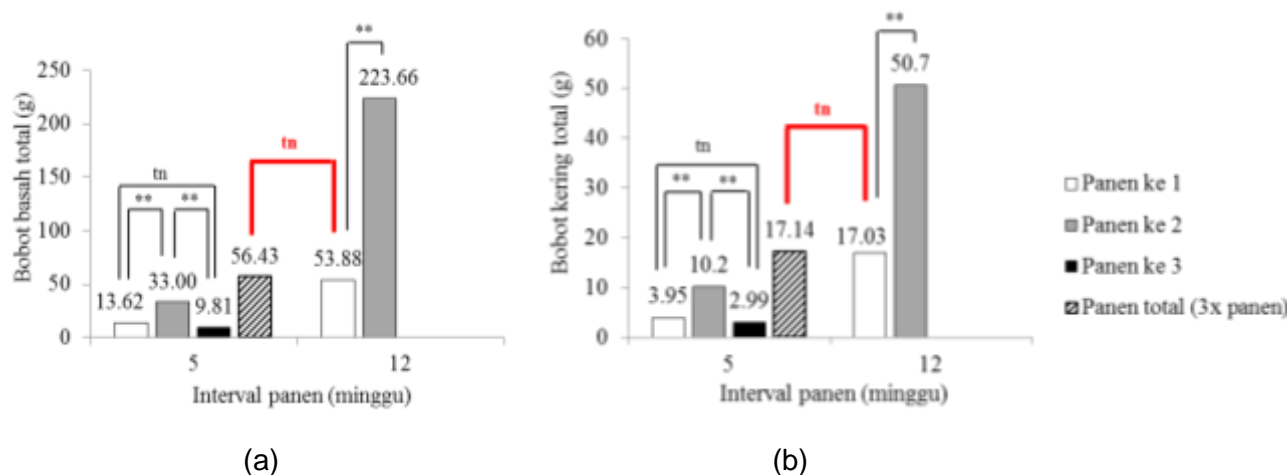


Figure 3. Total harvest fresh (a) and dry weight (b) of orange jessamine at 5 and 12-weeks harvest interval; \*\*: significantly different at  $P \leq 0.01$ , ns: not significant at  $P > 0.05$ .

The cumulative results of leaves dry weight, stem-branch dry weight and total dry weight of orange jessamine at the 5-week harvest interval in three harvests were not significantly different from the 12-week harvest interval (Figures 1b, 2b, and 3b). The dry weight increases linearly with a delay in harvest frequency (Kabi and Lutakome, 2013). Dry weight of orange jessamine decreases with increasing harvest frequency, this is because plants need time for the recovery phase that affects the recovery of carbohydrate reserves and decreases the level of dry matter production (Erdmann *et al.*, 1993).

The leaves dry weight to total dry weight proportion increased with increasing harvest frequency, while the proportion of stem-branch dry weight showed the opposite pattern. The percentage of leaves dry weight from the first harvest to the third harvest at the five-week harvest interval, respectively 69.87%, 78.24%, and 80.94%. Conversely, the percentage of stem-branch dry weight decreased in the next harvest at the five-week harvest interval, respectively 30.13%, 21.76% and 18.73%. The proportion of leaves dry weight in the 12-week harvest interval decreased from the first harvest to the second harvest, 87.84% and 71.62% respectively. Similar results also reported on cassava plants (Kang *et al.*, 2005).

Determination of crop intervals that are too short can have adverse effects on plants and ultimately affect the results produced. The monthly harvesting intervals of *Leucania*, *Gliricidia* and *Sesbania* plants which can increase mortality by 25% (Duguma *et al.*, 1988). Earlier leaves harvests and frequencies that more often reduce vegetative growth and yield of tuber seeds (*Xanthosoma sagittifolium*). This means that if the leaves are pruned earlier and more often, the strength of the source (leaves) decreases and causes photosynthate accumulation to the sink to decrease (Asumadu *et al.*, 2011).

#### Protein levels, PAL activity, phenolic levels, and total levels flavonoids

Protein is a polymer composed of several amino acids and is involved in growth, photosynthesis and homeostasis. Protein content in leaves has increased in the next harvest cycle, both at the five and 12-week harvest intervals (Figure 4a). Protein levels

are higher in *Desmanthus virgatus* plants which are more often harvested (Suksombat and Buakeeree, 2006). The results of other studies showing that leaves protein levels were influenced by harvest intervals also been reported in other plants (Amaglo *et al.*, 2006; Susanti *et al.*, 2011; Osadebe *et al.*, 2014)

Levels of active ingredients of medicinal plants vary according to the stages of growth and development of plants and parts of the organs analyzed, this is related to the activity of enzymes during the metabolic process of these active ingredients. The PAL activity of orange jessamine leaves has a pattern similar to protein levels in the five and 12-week harvest intervals (Figure 4b). It is presumed that the initial response from the openings (such as pruning) caused *de novo* synthesis and increased PAL activity (Campos-Vargas *et al.*, 2004). The treatment of the five-week harvest interval resulted in plants being gripped and causing an increase in PAL activity. This increase in PAL activity was not followed by an increase in flavonoid levels at the five-week harvest interval, while the increase in phenolic levels was not real. Phenolic levels in the first, second and third harvests were not significantly different at the five-week harvest interval, whereas at the twelve-week harvest interval the phenolic levels were significantly different between the first and second harvests (Figure 5a). Flavonoid levels at the five-week harvest interval had a pattern that was contrary to the levels of protein and PAL activity (Figures 4 and 5b). Flavonoid levels at the five-week harvest interval were highest in the first harvest cycle and decreased in the second and third harvest cycles, while at the twelve-week harvest interval flavonoid levels increased in the second harvest cycle. Decreased levels of flavonoids in the second harvest have also been reported in *Satureja hortensis*, *Majorana hortensis* and *Thymus vulgaris* (Vábková and Neugebauerová, 2011). This can be caused by other secondary metabolite products such as anthocyanins, and lignin which are synthesized during the change of phenolic compounds into flavonoid compounds. Therefore, the formation of flavonoids does not always depend on PAL activity (Cheng *et al.*, 2012).

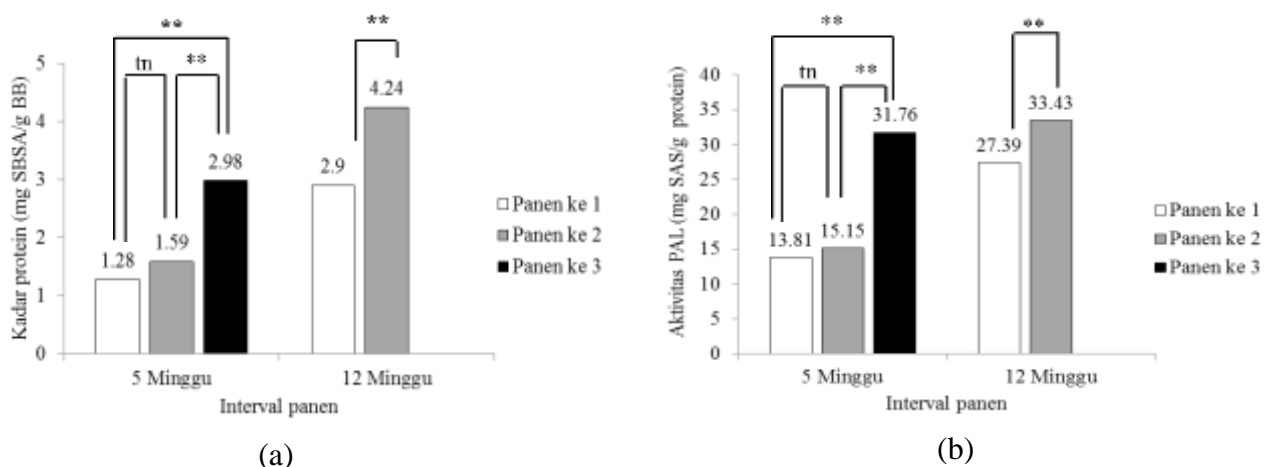


Figure 4. Protein content (a) and PAL activity (b) of orange jessamine at 5 and 12-weeks harvest interval; BSAE: bovine serum albumin equivalent, CAE: cinnamic acid equivalent, \*\*: significantly different



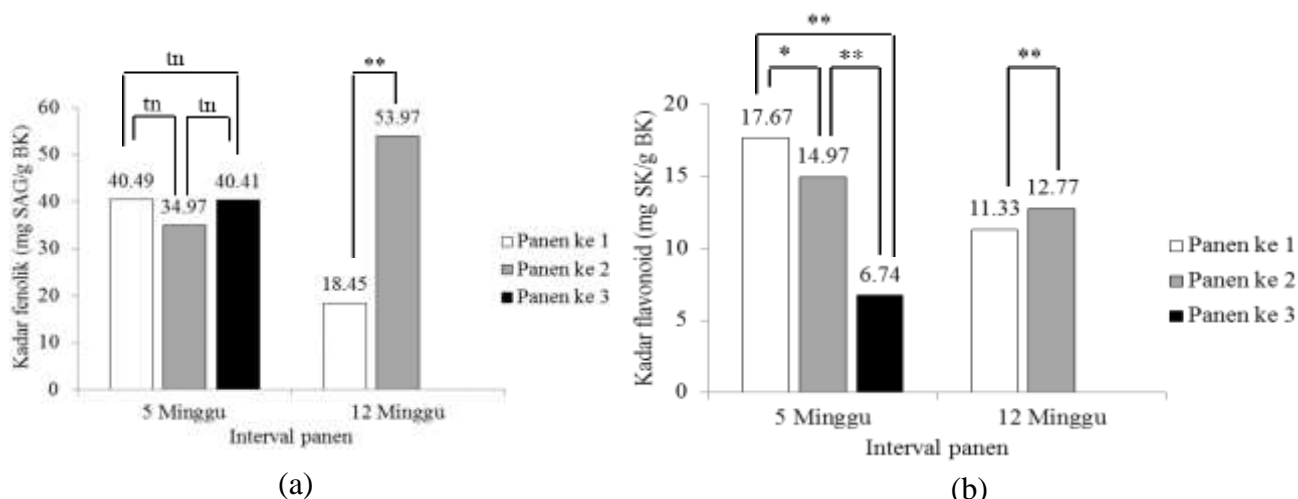


Figure 5. Phenolic (a) and flavonoid content (b) of orange jessamine at 5 and 12-weeks harvest interval; GAE: gallic acid equivalent, QE; quercetin equivalent, \*\*: significantly different at  $P \leq 0.01$ , ns: not significant at  $P > 0.05$ .

#### Metabolites production of per harvest cycle at two harvest intervals

Phenolic and flavonoid production in the second harvest at the five-week and twelve-week harvest intervals showed a marked increase, then decreased in the third harvest at the five-week harvest interval. Total leaves phenolic production of orange jessamine at the five-week harvest interval for three crops (0.48 g / plant) was not significantly different from the first harvest phenolic production at the twelve-week harvest interval (0.30 g / plant) (Figure 6). The total flavonoids production of orange jessamine leaves at the five-week harvest interval for three crops (0.18 g / plant) was also not significantly different from the production of the first harvest flavonoid at the twelve-week harvest interval (0.17 g / plant) (Figure 6).

The production of secondary metabolites is the result of multiplication of biomass with levels of secondary metabolites in it so that the value is determined by the harvested biomass and also the levels found at harvest. Increased production of active ingredients at the five-week harvest interval in the second harvest cycle is linear due to increased biomass. The production of active ingredients which decreased in the third harvest cycle at the five-week harvest interval was also caused by a decrease in biomass production. Short crop intervals in some plant species can suppress subsequent growth (Taylor *et al.*, 1979). In addition, the frequency of harvest and the stage of development of plants at harvest have a major influence on the quality of plant nutrients (Taylor *et al.*, 1977).

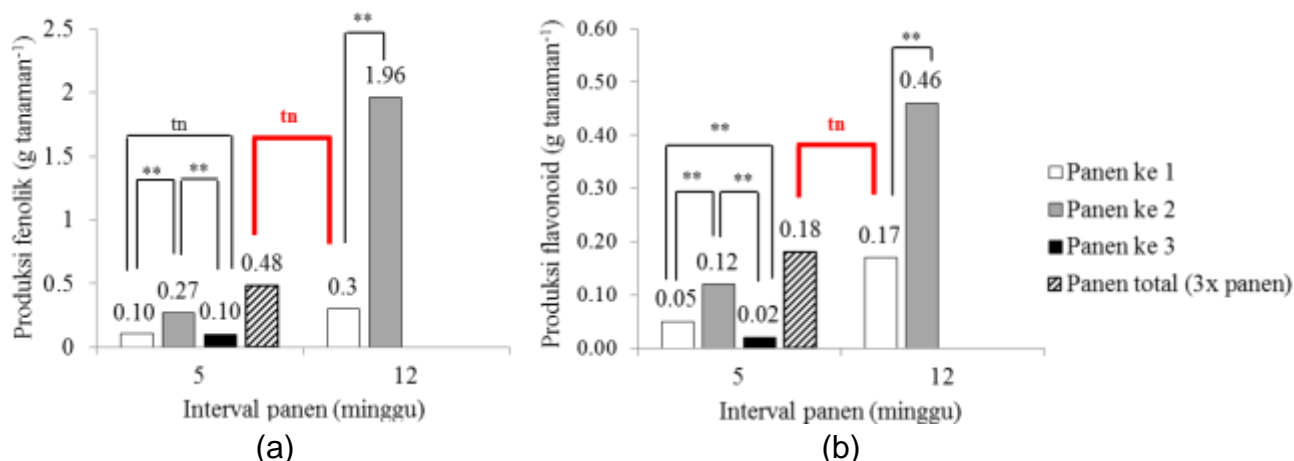


Figure 6. Phenolic and flavonoid production of orange jessamine leaves at 5 and 12-weeks harvest interval; \*\*: significantly different at  $P \leq 0.01$ , ns: not significant at  $P > 0.05$ .

#### 4. Conclusion

Proper harvesting arrangements are useful for obtaining optimal harvest biomass with maximum quality and ultimately increasing the efficacy of orange jessamine and maintaining plant growth so that production continuity can continue. The twelve-week harvest interval provides the best biomass and regrowth results for orange jessamine. The increase in PAL activity was not followed by an increase in phenolic and flavonoid levels at the five-week harvest interval. Phenolic and flavonoid production at the five-week harvest interval for three harvest cycles yielded no significant difference from the twelve-week harvest interval during one harvest cycle.

#### Acknowledgment

Thank you to the IPB Biopharmaca Research Center for funding this research. This research was conducted as part of the implementation of Strengthening and Efforts to Maintain the Sustainability of the Litbangrap Program at the Biopharmaca Research Center as the National Leading Center for the 2013 Fiscal Year.

#### References

- Amaglo N.K., Timpo G.M., Ellis W.O. and Bennet R.N. 2006. Effect of spacing and harvest frequency on the growth and leaf yield of moringa (*Moringa oleifera* Lam), a leafy vegetable crop. Proceedings of the International Workshop Moringa and other highly nutritious plant resources: strategies, standards and markets for a better impact on nutrition in Africa, Accra. Ghana, November 16-18:2006.[http://www.moringanews.org/doc/GB/Papers/Amaglo\\_Newton\\_text\\_G\\_B.pdf](http://www.moringanews.org/doc/GB/Papers/Amaglo_Newton_text_G_B.pdf).
- Asumadu H., Omenyo E.L. and Tetteh F. 2011. Physiological and economic implications of leaf harvesting on vegetative growth and cornel yield of cocoyam (*Xanthosoma sagittifolium*). Journal of Agronomy. 10(4): 112-117.

- Camm E.C. and Towers G.H.N. 1973. Phenylalanine ammonia lyase. *Phytochemistry*. 12: 961-973.
- Campos-Vargas R., Nonogaki H., Suslow T. and Saltveit M.E. 2004. Isolation and characterization of a wound inducible phenylalanine ammonia-lyase gene (LsPAL1) from Romaine lettuce leaves. *Physiologia Plantarum*. 121: 429-438.
- Chang C.C., Yang M.H., Wen H.M. and Chern J.C. 2002. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *Journal of Food and Drug Analysis*. 10(3): 178-182.
- Cheng S.Y., Xu F. and Wang Y. 2009. Advances in the study of flavonoids in *Ginkgo biloba* leaves. *Journal of Medicinal Plant Research*. 3(13): 1248-1252.
- Cheng S.Y., Xu F., Li L., Cheng H. and Zhang W. 2012. Seasonal pattern of flavonoid content and related enzyme activities in leaves of *Ginkgo biloba* L. *Notulae Botanicae Horti Agrobotanici*. 40(1): 98-106.
- Dangcham S., Bowen J., Ferguson I.B. and Ketsa S. 2008. Effect of temperature and low oxygen on pericarp hardening of mangosteen fruit stored at low temperature. *Postharvest Biology and Technology*. 50: 37-44.
- Duguma B., Kang B.T. and Okali D.U.U. 1988. Effect of pruning intensities of tree woody species grown in alley cropping with maize and cowpea on an alfisol. *Agroforestry Systems*. 6(1-3): 19-35.
- Erdmann T.K., Nair P.K.R. and Kang B.T. 1993. Effects of cutting height on reserve carbohydrates in *Gliricidia sepium* (Jacq.) Walp. *Forest Ecology and Management*. 57: 45-60.
- Gautam M.K., Singh A., Rao C.V. and Goel R.K. 2012a. Toxicological evaluation of *Murraya paniculata* (L.) leaves extract on rodents. *The American Journal of Pharmacology and Toxicology*. 7(2): 62-67.
- Gautam M.K., Gupta A., Vijaykumar M., Rao C.V. and Goel R.K. 2012b. Studies on the hypoglycemic effects of *Murraya paniculata* Linn. extract on alloxan-induced oxidative stress in diabetic and non diabetic models. *Asian Pacific Journal of Tropical Disease*. S186-S191.
- Gautam M.K., Anamika G., Rao C.V. and Goel R.K. 2012c. Antihyperglycemic and antioxidant potential of *Murraya paniculata* Linn. leaves: a preclinical study. *Journal of Pharmacy Research*. 5(3): 1334-1337.
- Iswantini D., Silitonga R.F., Martatilofa E. and Darusman L.K. 2011. Zingiber cassumunar, Guazuma ulmifolia, and Murraya paniculata extracts as antiobesity: In vitro: In vitro inhibitory effect on pancreatic lipase activity. *Hayati Journal of Biosciences*. 18(1): 6-10.
- Jones D.H. 1984. Phenylalanine ammonia-lyase: Regulation of its induction, and its role in plant development. *Phytochemistry*. 23(7): 1349-1359.
- Kabi F. and Lutakome P. 2013. Effect of harvesting *Gliricidia sepium* at different cutting frequencies on quantity and quality of herbage biomass for dairy cattle nutrition. *Journal of Animal Science Advances*. 3(6): 321-326.
- Kastono D., Sawitri H. dan Siswandono. 2005. Pengaruh nomor ruas setek dan dosis pupuk urea terhadap pertumbuhan dan hasil kumis kucing. *Ilmu Pertanian*. 12: 56-64.

- Khang D.N., Wiktorsson H. and Preston T.R. 2005. Yield and chemical composition of cassava foliage and tuber yield as influenced by harvesting height and cutting interval. *Asian-Australasian Journal of Animal Science*. 18(7): 1029-1035.
- Lowry O.H., Roseburgh N.J., Farr A.L and Randall R.J. 1951. Protein measurement with the folin phenol reagent. *The Journal of Biological Chemistry*. 193: 265-275.
- Lugasi A., Hóvari J., Sági K.V. and Biró L. 2003. The role of antioxidant phytonutrients in the prevention of diseases. *Acta Biologica Szegediensis*. 47(1-4): 119-125.
- Man N.V. and Wiktorsson H. 2003. Forage yield, nutritive value, feed intake and digestibility of three grass species as affected by harvest frequency. *Tropical Grassland*. 37(2): 101-110.
- Osadebe V.O., Echezona B.C. and Bakare S.O. 2014. Effect of weed management and cutting frequency on the leaf yield and proximate composition of fluted pumpkin (*Telfairia occidentalis* Hook F.). *American-Eurasian Journal of Agricultural & Environmental Sciences*. 14(7): 664-673.
- Pane M. 2010. Uji Efek Ekstrak Daun Kemuning (*Murraya paniculata* (L.) Jack) sebagai Penurun Kadar Kolesterol Darah Marmut Jantan (*Cavia cobaya*). Skripsi. Medan(ID): Universitas Sumatera Utara.
- Paramaguru R., Janaki P.S., Eswaran M.B., Rao C.V., Rawat A.K.S and Vijayakumar M. 2012. Free radical scavenging and lipid peroxidation inhibition potential of various fractions of *Murraya paniculata*. *Pharmacologia* 3(5): 138-143.
- Podder M.K., Das B.N., Saha A. and Ahmed M. 2011. Analgesic activity of bark of *Murraya paniculata*. *International Journal of Medicinal Sciences*. 3(4): 105-108.
- Pourmorad F., Hosseinimehr S.J. and Shahabimajd N. 2006. Antioxidant activity, phenol, and flavonoid contents of some selected Iranian medical plants. *African Journal of Biotechnology*. 5(11): 1142-1145.
- Rahman A., Hasanuzzaman, Uddin N. and Shahid I.Z. 2010. Antidiarrhoeal and anti-inflammatory activities of *Murraya paniculata* (L.) Jack. *Pharmacologyonline*. #: 768-776.
- Sajimin N.D., Purwantari, Sutedi E. dan Oyo. 2011. Pengaruh interval potong terhadap produktivitas dan kualitas tanaman bangun-bangun (*Coleus amboinicus* L.) sebagai komoditas harapan pakan ternak. *Jurnal Ilmu Ternak dan Veteriner*. 16(4): 288-293.
- Suksombat W. and Buakeeree K. 2006. Effect of cutting interval and cutting height on yield and chemical composition of hedge lucerne (*Desmanthus virgatus*). *Asian-Australasian Journal of Animal Sciences*. 19(1): 31-34.
- Susanti H., Aziz S.A., Melati M. and Susanto S. 2011. Protein and anthocyanin production of waterleaf shoots (*Talinum triangulare* (Jacq.) Willd) at different levels of nitrogen+potassium and harvest intervals. *Jurnal Agronomi Indonesia*. 39(2): 119-123.
- Syahadat R.M. and Aziz S.A. 2012. Pengaruh komposisi media dan fertigasi pupuk organik terhadap kandungan bioaktif daun tanaman kemuning di pembibitan. *Buletin Penelitian Tanaman Rempah dan Obat*. 23(2): 142-147.

- Taylor A.O., Hughes K.A., Hunt B.J. and Latch G.C.M. 1979. Annual cool season legumes for forage. I. A survey of lines for yield and disease resistance at Kaitaia and Palmerston North. N. Z. J. Exp. Agric. 7: 45-49.
- Taylor A.O., Hughes K.A., Haslemore R.M. and Holland R. 1977. Influence of maturity and frequency of harvest on the nutritive quality of cool season forage legumes. Ibid. 7: 45-49.
- Tapiero H., Tew K.D., Nguyen B.G. and Mathe G. 2002. Polyphenols: do they play a role in the prevention of human pathologies?. Biomedicine and Pharmacotherapy. 56: 200-207.
- Vábková J. and Neugebauerová J. 2011. Determination of total phenolic content, total flavonoid content and FRAP in culinary herbs in relation to harvest time. Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis. LX (1): 167-171.
- Ververidis F., Trantas E., Douglas C., Vollmer G., Kretzschmar G. and Panopoulos N. 2007. Biotechnology of flavonoids and other phenylpropanoid-derived natural products. Biotechnology Journal. 2(10): 1214-1234.
- Waterhouse L.A. 2002. Determination of Total Phenolics: Current Protocols in Food Analytical Chemistry. John Wiley & Sons, Inc., USA.
- Wu L., Li P., Wang X., Zhuang Z., Farzaneh F. and Xu R. 2010. Evaluation of anti-inflammatory and antinociceptive activities of *Murraya exotica*. Pharmaceutical Biology. 48: 1344-1353.
- Xiao P.G. and Wang N.G. 1991. Can ethnopharmacology contribute to the development of anti-fertility drugs. Journal of Ethnopharmacology. 32: 167-77.