

Research Article

Supervised Clustering Based on DPCLUSO: Prediction of Plant-Disease Relations Using Jamu Formulas of KNAPSAcK Database

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Indonesia has the largest medicinal plant species in the world and these plants are used as Jamu medicines. Jamu medicines are popular traditional medicines from Indonesia and we need to systemize the formulation of Jamu and develop basic scientific principles of Jamu to meet the requirement of Indonesian Healthcare System. We propose a new approach to predict the relation between plant and disease using network analysis and supervised clustering. At the preliminary step, we assigned 3138 Jamu formulas to 116 diseases of International Classification of Diseases (ver. 10) which belong to 18 classes of disease from National Center for Biotechnology Information. The correlation measures between Jamu pairs were determined based on their ingredient similarity. Networks are constructed and analyzed by selecting highly correlated Jamu pairs. Clusters were then generated by using the network clustering algorithm DPCLUSO. By using matching score of a cluster, the dominant disease and high frequency plant associated to the cluster are determined. The plant to disease relations predicted by our method were evaluated in the context of previously published results and were found to produce around 90% successful predictions.

1. Introduction

Big data biology, which is a discipline of data-intensive science, has emerged because of the rapid increasing of data in omics fields such as genomics, transcriptomics, proteomics, and metabolomics as well as in several other fields such as ethnomedicinal survey. The number of medicinal plants is estimated to be 40,000 to 70,000 around the world [1] and many countries utilize these plants as blended herbal medicines, for example, China (traditional Chinese medicine), Japan (Kampo medicine), India (Ayurveda, Sidha, and Unani), and Indonesia (Jamu). Nowadays, the use

of traditional medicines is rapidly increasing [2, 3]. These medicines consist of ingredients made from plants, animals, minerals, or combination of them. The traditional medicines have been used for generations for treatments of diseases or maintaining health of people and the most popular form of traditional medicine is herbal medicine. Blended herbal medicines as well as single herb medicines include a large number of constituent substances which exert effects on human physiology through a variety of biological pathways. The KNAPSAcK Family database systems can be used to comprehensively understand the medicinal usage of plants based upon traditional and modern knowledge [4, 5]. This

TABLE 1: List of diseases using International Classification of Diseases ver. 10 (class of disease IDs correspond to Table 2).

ID	Disease	Class of disease
1	Abdominal pain	3
2	Abdominal pain, diarrhea	3
3	Acne	16
4	Acne, skin problems (cosmetics)	16
5	Amenorrhoea, dysmenorrhoea	6
6	Amenorrhoea, irregular menstruation	6
7	Anaemia	1
8	Appendicitis, urinary tract infection, tonsillitis	3
9	Arthralgia	11
10	Arthralgia, arthritis	11
11	Asthma	15
12	Benign prostatic hyperplasia (Bph)	10
13	Breast disorder	6
14	Bromhidrosis	16
15	Bronchitis	15
16	Cancer	2
17	Cancer pain	2
18	Cancer, inflammation	2
19	Colic abdomen, bloating (in infant)	3
20	Common cold	15
21	Common cold, dyspepsia, insect bites	15, 3, 16
22	Common cold, influenza	15
23	Cough	15
24	Degenerative disease	14
25	Dermatitis, urticaria, erythema	16
26	Diabetes	14
27	Diabetic gangrene	16
28	Diarrhea	3
29	Diarrhea, abdominal pain	3
30	Diseases of the eye	5
31	Disorders in pregnancy	6
32	Dysmenorrhea	6
33	Dysmenorrhea, irregular menstruation	6
34	Dysmenorrhea, menstrual syndrome	6
35	Dyspepsia	3
36	Dyspnoea	15
37	Dyspnoea, cough, orthopnoea	15
38	Fatigue	11
39	Fatigue, anaemia, loss appetite	1
40	Fatigue, lack of sexual function	6
41	Fatigue, low back pain	11
42	Fatigue, myalgia, arthralgia	11
43	Fatigue, osteoarthritis	11
44	Fertility problem	6, 10
45	Fever	0

TABLE 1: Continued.

ID	Disease	Class of disease
46	Gastritis, gastric ulcer	3
47	Haemorrhoids	1
48	Headache	13
49	Heart diseases	8
50	Heartburn	3, 8
51	Hepatitis, other diseases of liver	3
52	Hypercholesterolaemia	14
53	Hypertension	8
54	Hypertension, diabetes	14
55	Hypertension, hypercholesterolaemia	14
56	Hyperuricemia	1
57	Immunodeficiency	9
58	Indigestion (K.30)	3
59	Indigestion, lose appetite	3
60	Infertility	6, 10
61	Irregular menstruation, menstruation syndrome	6
62	Kidney diseases	17
63	Lactation problems	6
64	Leukorrhoea (Vaginalis)	6
65	Leukorrhoea (Vaginalis), dysmenorrhoea	6
66	Lose appetite	3
67	Lose appetite, underweight	14
68	Low back pain, myalgia, arthralgia	11
69	Low back pain, myalgia, constipation	11
70	Low back pain, urinary tract infection	17
71	Lung diseases	15
72	Malaise and Fatigue	11
73	Malaise and Fatigue, Constipation	11
74	Malaise and Fatigue, Fertility Problems	10, 11
75	Malaise and Fatigue, Low Back Pain	11
76	Malaise and Fatigue, Sexual Dysfunction	11, 6, 10
77	Malaise and Fatigue, Skin Problems (Cosmetics)	16
78	Malaria, anaemia	1
79	Meno-metrorrhagia	6
80	Menopausal syndrome	6
81	Menopause/menstrual syndrome, leukorrhoea (vaginalis)	6
82	Menstrual syndrome	6
83	Menstrual syndrome, fatigue	6
84	Migraine	13
85	Mood disorder	18
86	Myalgia, arthralgia	11
87	Nausea/vomiting of pregnancy	6
88	Osteoarthritis	11
89	Osteoarthritis, fatigue	11

TABLE 1: Continued.

ID	Disease	Class of disease
90	Overweight, obesity	14
91	Paralysis	13
92	Post partum syndrome	6
93	Prevent from overweight	14
94	Respiratory infection due to smoking	15
95	Respiratory tract infection	15
96	Rheumatoid arthritis, gout	11
97	Secondary amenorrhea	6
98	Secondary amenorrhea, irregular menstruation	6
99	Sexual dysfunction, fatigue	6, 10
100	Skin diseases	16
101	Skin problems (cosmetics)	16
102	Sleeping and Mood Disorders	18
103	Sleeping disorders	18
104	Stomatitis	3
105	Stomatitis, gingivitis, tonsilitis	3
106	Stone in kidney (N20.0)	17
107	Stone in kidney (N20.0), urinary bladder stone (N21.0)	17
108	Tonsilitis	4
109	Tonsilofaringitis	4
110	Toothache	13
111	Typhoid, dyspepsia	3
112	Ulcer of anus and rectum	3
113	Underweight, lose appetite	3
114	Urinary tract infection (urethritis)	17
115	Vaginal discharges	6
116	Vaginal diseases	6

database has information about the selected herbal ingredients, that is, the formulas of Kampo and Jamu, omics information of plants and humans, and physiological activities in humans. Jamu is generally composed based on the experience of the users for decades or even hundreds of years. However, versatile scientific analyses are needed to support their efficacy and their safety. Attaining this objective is in accordance with the 2010 policy of the Ministry of Health of Indonesian Government about scientification of Jamu. Thus, it is required to systemize the formulations and develop basic scientific principles of Jamu to meet the requirement of Indonesian Healthcare System. Afendi et al. initiated and conducted scientific analysis of Jamu for finding the correlation between plants, Jamu, and their efficacy using statistical methods [6–8]. They used Biplot, partial least squares (PLS), and bootstrapping methods to summarize the data and also focused on prediction of Jamu formulations. These methods give a good understanding about relationship between plants, Jamu, and their efficacy. Among 465 plants used in 3138 Jamu, 190 plants were shown to be effective for at least one efficacy and these plants were considered

to be the main ingredients of Jamu. The other 275 plants are considered to be supporting ingredients in Jamu because their efficacy has not been established yet.

Network biology can be defined as the study of the network representations of molecular interactions, both to analyze such networks and to use them as a tool to make biological predictions [9]. This study includes modelling, analysis, and visualizations, which holds important task in life science today [10]. Network analysis has been increasingly utilized in interpreting high throughput data on omics information, including transcriptional regulatory networks [11], coexpression networks [12], and protein-protein interactions [13]. We can easily describe relationship between entities in the network and also concentrate on part of the network consisting of important nodes or edges. These advantages can be adopted for analyzing medicinal usage of plants in Jamu and diseases. Network analysis provides information about groups of Jamu that are closely related to each other in terms of ingredient similarity and thus allows precise investigation to relate plants to diseases. On the other hand, multivariate statistical methods such as PLS can assign plants to efficacy by global linear modeling of the Jamu ingredients and efficacy. However, there is still lack of appropriate network based methods to learn how and why many plants are grouped in certain Jamu formula and the combination rule embedding numerous Jamu formulas.

It is needed to explore the relationship between Indonesian herbal plants used in Jamu medicines and the diseases which are treated using Jamu medicines. When effectiveness of a plant against a disease is firmly established, then further analysis about that plant can be proceeded to molecular level to pinpoint the drug targets. The present study developed a network based approach for prediction of plant-disease relations. We utilized the Jamu data from the KNAPSAcK database. A Jamu network was constructed based on the similarity of their ingredients and then Jamu clusters were generated using the network clustering algorithm DPCLUSO [14, 15]. Plant-disease relations were then predicted by determining the dominant diseases and plants associated with selected Jamu clusters.

2. Methods

2.1. Concept of the Methodology. Jamu medicines consist of combination of medicinal plants and are used to treat versatile diseases. In this work we exploit the ingredient similarity between Jamu medicines to predict plant-disease relations. The concept of the proposed method is depicted in Figure 1. In step 1 a network is constructed where a node is a Jamu medicine and an edge represents high ingredient similarity between the corresponding Jamu pair. In Figure 1, the nodes of the same color indicate the Jamu medicines used for the same disease. The similarity is represented by Pearson correlation coefficient [16, 17]; that is,

$$\text{corr}(X, Y) = \frac{\sum_{i=1}^l (x_i - \bar{x})(y_i - \bar{y})}{\sqrt{\sum_{i=1}^l (x_i - \bar{x})^2 \sum_{i=1}^l (y_i - \bar{y})^2}}, \quad (1)$$

TABLE 2: Distribution of Jamu formulas according to 18 classes of disease (classes of diseases are determined by NCBI in ID1 to ID16 and by the present study in ID17 and ID18 represented by asterisks in Ref. columns).

ID	Class of disease (NCBI)	Ref.	Number of Jamu	Percentage
1	Blood and lymph diseases	NCBI	201	6.41
2	Cancers	NCBI	32	1.02
3	The digestive system	NCBI	457	14.56
4	Ear, nose, and throat	NCBI	2	0.06
5	Diseases of the eye	NCBI	1	0.03
6	Female-specific diseases	NCBI	382	12.17
7	Glands and hormones	NCBI	0	—
8	The heart and blood vessels	NCBI	57	1.82
9	Diseases of the immune system	NCBI	22	0.70
10	Male-specific diseases	NCBI	17	0.54
11	Muscle and bone	NCBI	649	20.68
12	Neonatal diseases	NCBI	0	—
13	The nervous system	NCBI	32	1.02
14	Nutritional and metabolic diseases	NCBI	576	18.36
15	Respiratory diseases	NCBI	313	9.97
16	Skin and connective tissue	NCBI	163	5.19
17	The urinary system	*	90	2.87
18	Mental and behavioral disorders	*	21	0.67
The number of Jamu classified into multiple disease classes			119	3.79
The number of Jamu unclassified			4	0.13
Total Jamu formulas			3138	100.00

where x_i is the weight of plant- i in Jamu X , y_i is the weight of plant- i in Jamu Y , \bar{x} is mean of Jamu X , and \bar{y} is mean of Jamu Y . The higher similarity between Jamu pairs the higher the correlation value. In the present study, x_i and y_i are assigned as 1 or 0 in cases the i th plant is, respectively, included or not included in the formula. Under such condition, Pearson correlation corresponds to fourfold point correlation coefficient; that is,

$$\text{corr}(X, Y) = \frac{ad - bc}{\sqrt{(a+b)(a+c)(b+d)(c+d)}}, \quad (2)$$

where a , b , c , and d represent the numbers of plants included in both X and Y , in only X , in only Y , and in neither X nor Y , respectively.

In step 2 the Jamu clusters are generated using network clustering algorithm DPCLUSO. DPCLUSO can generate clusters characterized by high density and identified by periphery; that is, the Jamu medicines belonging to a cluster are highly cohesive and separated by a natural boundary. Such clusters contain potential information about plant-disease relations.

In step 3 we assess disease-dominant clusters based on matching score represented by the following equation:

$$\begin{aligned} &\text{matching score} \\ &= \frac{\text{number of Jamu belonging to the same disease}}{\text{total number of Jamu in the cluster}}. \end{aligned} \quad (3)$$

Matching score of a cluster is the ratio of the highest number of Jamu associated with a single disease to the total number of Jamu in the cluster. We assign a disease to a cluster for which the matching score is greater than a threshold value. In step 4, we determine the frequency of plants associated with a cluster if and only if a disease is assigned to it in the previous step. The highest frequency plant associated to a cluster is considered to be related to the disease assigned to that cluster. True positive rates (TPR) or sensitivity was used to evaluate resulting plants. TPR is the proportion of the true positive predictions out of all the true predictions, defined by the following formula [18]:

$$\text{TPR} = \frac{\text{TP}}{\text{TP} + \text{FN}}, \quad (4)$$

where true positive (TP) is the number of correctly classified and false negative (FN) is the number of incorrectly rejected entities. We refer to the proposed method as supervised clustering because after generation of the clusters we narrow down the candidate clusters for further analysis based on supervised learning and thus improve the accuracy of prediction of the proposed method.

3. Result and Discussion

3.1. Construction and Comparison of Jamu and Random Networks. We used the same number of Jamu formulas from previous research [6], 3138 Jamu formulas, and the set union

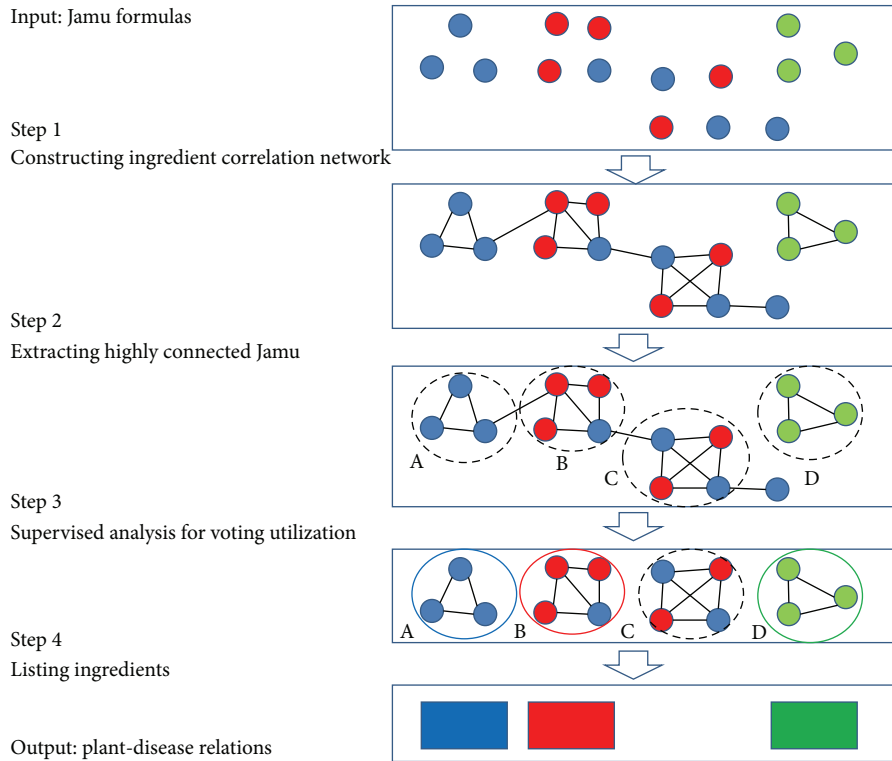


FIGURE 1: Concept of the methodology: network construction based on ingredient similarity between individual Jamu medicines, network clustering, and classification of medicinal plants to dominant disease.

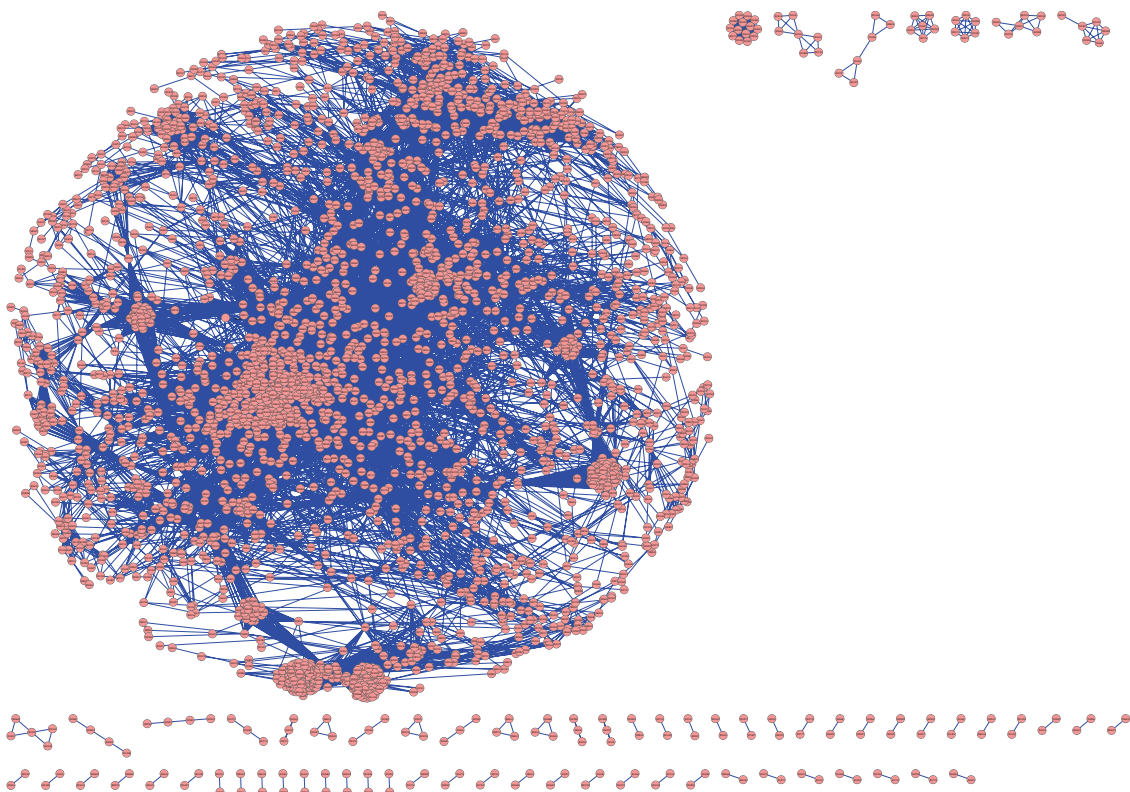


FIGURE 2: The network consisting of 0.7% Jamu pairs (correlation value above or equal to 0.596).

TABLE 3: Statistics of three datasets.

	Parameters	0.7%	0.5%	0.3%
Network statistics	Total pairs	34,454	24,610	14,766
	Minimum correlation	0.596	0.665	0.718
	Number of Jamu formulas	2,779	2,496	2,085
	Average degree	24.8	19.7	14.2
	(Random network: ER)	(24.8 ± 0.0)	(19.7 ± 0.0)	(14.2 ± 0.0)
	(Random network: BA)	(24.7 ± 0.1)	(19.7 ± 0.1)	(14.1 ± 0.1)
	(Random network: CNN)	(24.7 ± 0.4)	(19.7 ± 0.4)	(14.0 ± 0.4)
	Clustering coefficient	0.521	0.520	0.540
	(Random network: ER)	(0.009 ± 0.000)	(0.008 ± 0.000)	(0.007 ± 0.000)
	(Random network: BA)	(0.030 ± 0.001)	(0.028 ± 0.001)	(0.026 ± 0.001)
	(Random network: CNN)	(0.246 ± 0.008)	(0.239 ± 0.008)	(0.233 ± 0.010)
	Number of connected components	69	119	254
	(Random networks: ER, BA, CNN)	(1)	(1)	(1)
	Network diameter	15	17	20
	(Random network: ER)	(4.0 ± 0.0)	(4.0 ± 0.0)	(5.0 ± 0.0)
	(Random network: BA)	(10.8 ± 0.8)	(11.2 ± 1.5)	(10.8 ± 0.9)
	(Random network: CNN)	(14.6 ± 1.9)	(14.1 ± 1.4)	(14.7 ± 1.3)
	Network density	0.008	0.008	0.007
	(Random network: ER)	(0.009 ± 0.000)	(0.008 ± 0.000)	(0.007 ± 0.000)
	(Random network: BA)	(0.009 ± 0.000)	(0.008 ± 0.000)	(0.007 ± 0.000)
(Random network: CNN)	(0.009 ± 0.000)	(0.008 ± 0.000)	(0.007 ± 0.000)	
DPCLUSO	Total number of clusters	1,746	1,411	938
	Number of clusters with more than 2 Jamu (%)	1,296 (74.2)	873 (61.9)	453 (48.3)
	Number of Jamu formulas in the biggest cluster	118	104	89

of all formulas consists of 465 plants. We assigned 3138 Jamu formulas to 116 diseases of International Classification of Diseases (ICD) version 10 from World Health Organization (WHO, Table 1) [19]. Those 116 diseases are mapped to 18 classes of disease, which contains 16 classes of disease from National Center for Biotechnology Information (NCBI) [20] and 2 additional classes. Table 2 shows distribution of 3138 Jamu into 18 classes of disease. According to this classification, most Jamu formulas are useful for relieving muscle and bone, nutritional and metabolic diseases, and the digestive system. Furthermore, there is no Jamu formula classified into glands and hormones and neonatal disease classes. We excluded 4 Jamu formulas which are used to treat fever in the evaluation process because this symptom is very general and almost appeared in all disease classes. Jamu-plant-disease relations can be represented using 2 matrices: first matrix is Jamu-plant relation with dimension 3138×465 and the second matrix is Jamu-disease relation with dimension 3138×18 .

After completion of data acquisition process, we calculated the similarity between Jamu pairs using correlation measure. The similarity measures between Jamu pairs were determined based on their ingredients. Corresponding to K (3138 in present case) Jamu formulas, there can be maximum $(K \times (K - 1)/2) = (3138 \times (3137/2)) = 4,921,953$ Jamu

pairs. We sorted the Jamu pairs based on correlation value using descending order and selected top- n (0.7%, 0.5%, and 0.3%) pairs of Jamu formula to create 3 sets of Jamu pairs. The number of Jamu pairs for 0.7%, 0.5%, and 0.3% datasets is 34,454 pairs, 24,610 pairs, and 14,766 pairs and the corresponding minimum correlation values are 0.596, 0.665, and 0.718, respectively. The three datasets of Jamu pairs can be regarded as three undirected networks (step 1 in Figure 1) consisting of 2779, 2496, and 2085 Jamu formulas, respectively (Table 3). Figure 2 shows visualization of 0.7% Jamu networks using Cytoscape Spring Embedded layout. We verified that the degree distributions of the Jamu networks are somehow close to those of scale-free networks, that is, roughly are of power law type. However, in the high-degree region the power law structure is broken (Figure 3). Nearly accurate relation of power laws between medicinal herbs and the number of formulas utilizing them was observed in Jamu system but not in Kampo (Japanese crude drug system) [4]. The difference of formulas between Jamu and Kampo can be explained by herb selection by medicinal researchers based on the optimization process of selection [4]. Thus, the broken structure of power law corresponding to Jamu networks is associated with the fact that selection of Jamu pairs based on ingredient correlation leads to nonrandom selection. We also constructed random networks according

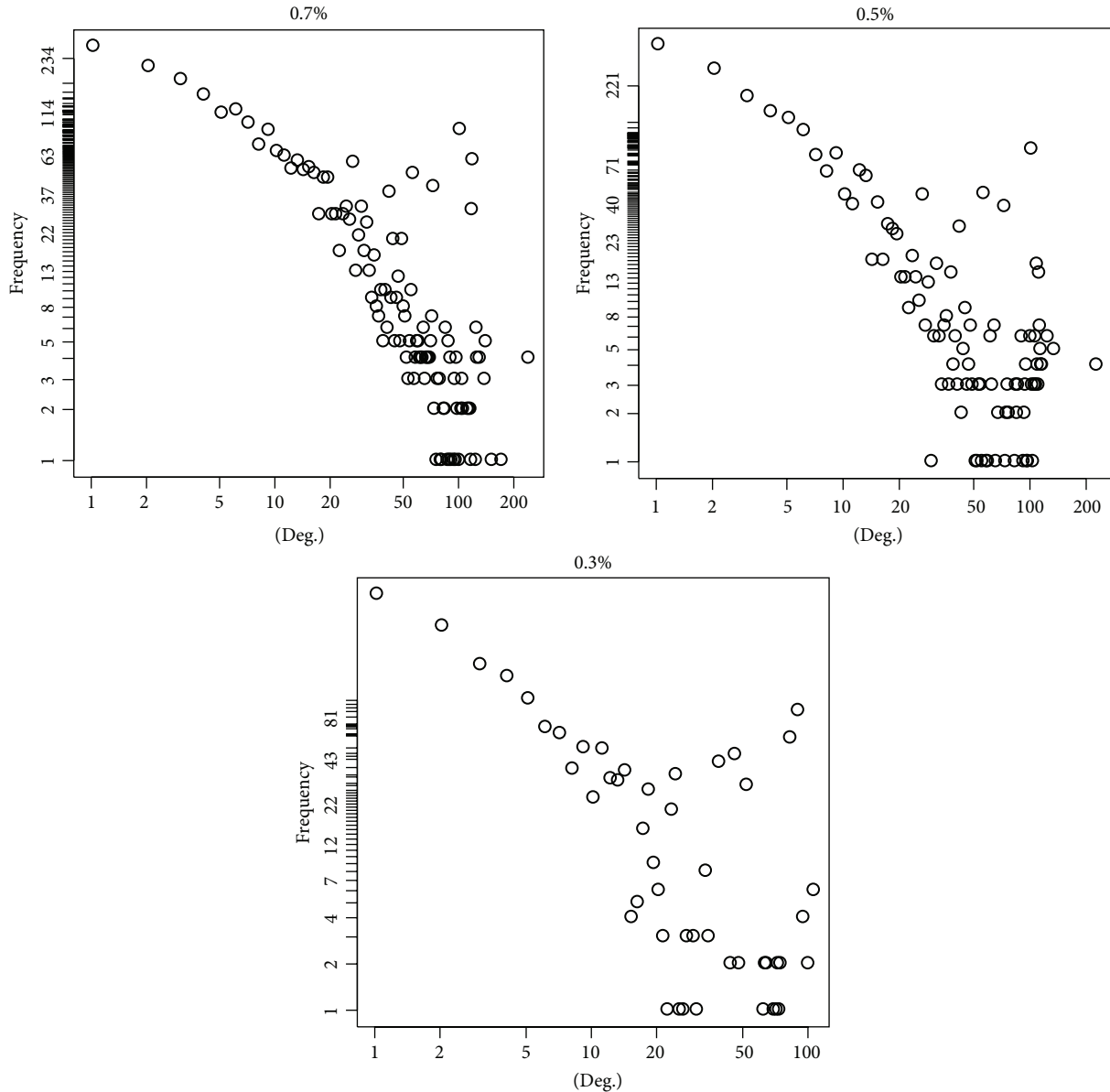


FIGURE 3: Degree distributions of three Jamu networks roughly follow power law. The x -axis corresponds to the log of degree of a node in the Jamu network and the y -axis corresponds to the log of the number of Jamu.

to Erdős-Rényi (ER) model [21], Barabási-Albert (BA) model [22], and Vazquez’s Connecting Nearest Neighbor (CNN) model [23] of the same size corresponding to each of the real Jamu network. We used Cytoscape Network Analyzer plugin [24] and R software for analyzing the characteristics of both the Jamu and the random networks.

We determined five statistical indexes, that is, average degree, clustering coefficient, number of connected component, network diameter, and network density of each Jamu network and also of each random network. The clustering coefficient C_n of a node n is defined as $C_n = 2e_n / (k_n(k_n - 1))$, where k_n is the number of neighbors of n and e_n is the number of connected pairs between all neighbors of n . The network diameter is the largest distance between any two nodes. If

a network is disconnected, its diameter is the maximum of all diameters of its connected components. A network’s density is the ratio of the number of edges in the network over the total number of possible edges between all pairs of nodes (which is $n(n - 1)/2$, where n is the number of vertices, for an undirected graph). The average number of neighbors and the network density are the same for the real and random networks of the same size as it is shown in Table 3. In case of 0.7% and 0.5% real networks, the clustering coefficient is roughly the same and in case of 0.3% the clustering coefficient is somewhat larger. The number of connected components and the diameter of the Jamu networks gradually decrease as the network grows bigger by addition of more nodes and edges.

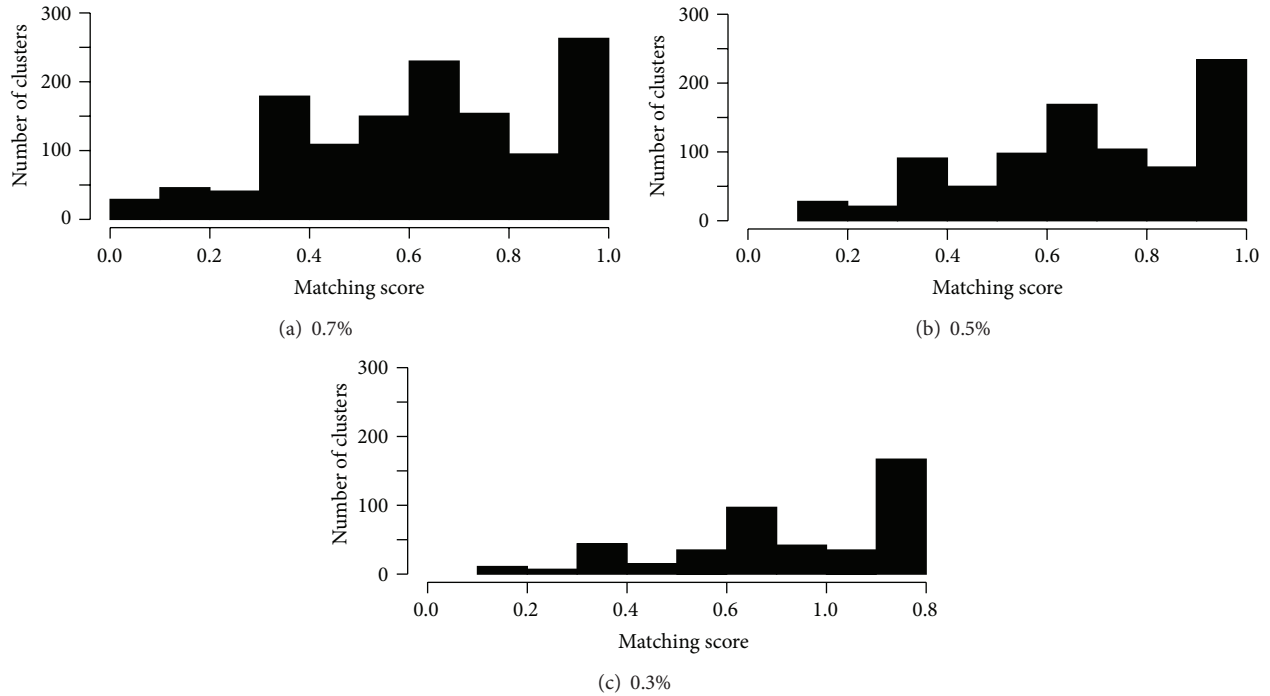


FIGURE 4: Distribution of clusters based on matching score.

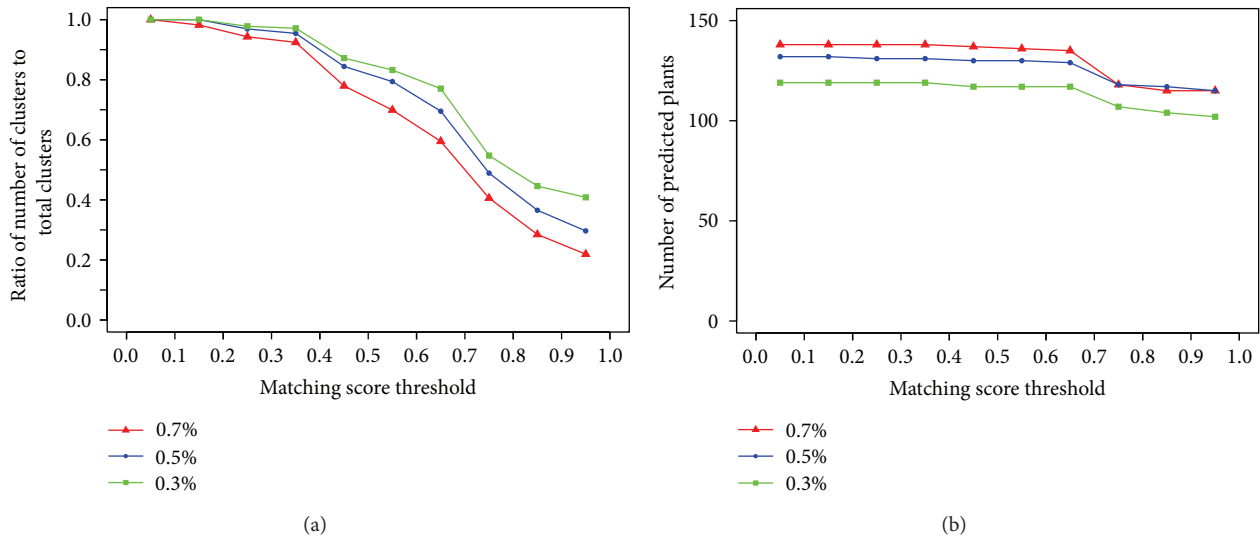


FIGURE 5: (a) Success rate and (b) number of predicted plants with respect to matching score thresholds.

Very different values corresponding to clustering coefficient, connected component, and network diameter imply that the Jamu networks are quite different from all 3 types of random networks. The differences between Jamu networks and ER random networks are the largest. Random networks constructed based on other two models are also substantially different from Jamu networks. Based on the fact that the random networks constructed based on all three types of models are different from the Jamu networks, it can be concluded that structure of Jamu networks is reasonably biased and thus might contain certain information about

plant-disease relations. Specially, much higher value corresponding to clustering coefficient indicates that there are clusters in the networks worthy to be investigated. To extract clusters from the Jamu networks (step 2 in Figure 1) we applied DPCLUSO network clustering algorithm [14] to generate overlapping clusters based on density and periphery tracking.

3.2. Supervised Clustering Based on DPCLUSO. DPCLUSO is a general-purpose clustering algorithm and useful for finding overlapping cohesive groups in an undirected simple graph

TABLE 4: List of plants assigned to each disease.

Number	Plants name	Hit-miss status	
A. Disease: blood and lymph diseases			
1	<i>Tamarindus indica</i>	Hit	*
2	<i>Allium sativum</i>	Hit	*
3	<i>Tinospora tuberculata</i>	Hit	*
4	<i>Piper retrofractum</i>	Hit	
5	<i>Syzygium aromaticum</i>	Hit	*
6	<i>Bupleurum falcatum</i>	Hit	
7	<i>Graptophyllum pictum</i>	Hit	
8	<i>Plantago major</i>	Hit	
9	<i>Zingiber officinale</i>	Hit	*
10	<i>Cinnamomum burmannii</i>	Hit	*
11	<i>Soya max</i>	Miss	*
12	<i>Kaempferia galanga</i>	Hit	
13	<i>Curcuma longa</i>	Hit	*
14	<i>Piper nigrum</i>	Hit	
15	<i>Zingiber aromaticum</i>	Hit	*
16	<i>Phyllanthus urinaria</i>	Hit	*
17	<i>Oryza sativa</i>	Hit	
18	<i>Myristica fragrans</i>	Hit	*
19	<i>Alstonia scholaris</i>	Hit	*
20	<i>Syzygium polyanthum</i>	Miss	
21	<i>Andrographis paniculata</i>	Hit	*
22	<i>Sida rhombifolia</i>	Miss	
23	<i>Cyperus rotundus</i>	Hit	
24	<i>Sonchus arvensis</i>	Miss	
25	<i>Curcuma aeruginosa</i>	Hit	*
26	<i>Curcuma xanthorrhiza</i>	Hit	
B. Disease: cancers			
1	<i>Catharanthus roseus</i>	Hit	
C. Disease: the digestive system			
1	<i>Foeniculum vulgare</i>	Hit	
2	<i>Glycyrrhiza uralensis</i>	Hit	*
3	<i>Imperata cylindrica</i>	Hit	
4	<i>Zingiber purpureum</i>	Hit	*
5	<i>Physalis peruviana</i>	Hit	
6	<i>Punica granatum</i>	Hit	*
7	<i>Echinacea purpurea</i>	Hit	
8	<i>Zingiber officinale</i>	Hit	*
9	<i>Psidium guajava</i>	Hit	
10	<i>Baeckea frutescens</i>	Hit	*
11	<i>Amomum compactum</i>	Hit	
12	<i>Cinnamomum burmannii</i>	Hit	*
13	<i>Melaleuca leucadendra</i>	Hit	
14	<i>Caesalpinia sappan</i>	Hit	*
15	<i>Parkia roxburghii</i>	Hit	
16	<i>Rheum tanguticum</i>	Hit	
17	<i>Kaempferia galanga</i>	Hit	
18	<i>Coriandrum sativum</i>	Hit	

TABLE 4: Continued.

Number	Plants name	Hit-miss status	
19	<i>Curcuma longa</i>	Hit	
20	<i>Zingiber aromaticum</i>	Hit	
21	<i>Phyllanthus urinaria</i>	Hit	
22	<i>Myristica fragrans</i>	Hit	
23	<i>Hydrocotyle asiatica</i>	Hit	*
24	<i>Carica papaya</i>	Hit	
25	<i>Mentha arvensis</i>	Hit	
26	<i>Lepiniopsis ternatensis</i>	Hit	
27	<i>Helicteres isora</i>	Hit	
28	<i>Andrographis paniculata</i>	Hit	
29	<i>Symplocos odoratissima</i>	Hit	
30	<i>Schisandra chinensis</i>	Hit	
31	<i>Blumea balsamifera</i>	Hit	
32	<i>Silybum marianum</i>	Hit	*
33	<i>Cinnamomum sintoc</i>	Hit	
34	<i>Elephantopus scaber</i>	Hit	
35	<i>Curcuma aeruginosa</i>	Hit	
36	<i>Kaempferia pandurata</i>	Hit	
37	<i>Curcuma xanthorrhiza</i>	Hit	
38	<i>Curcuma mangga</i>	Hit	*
39	<i>Curcuma zedoaria</i>	Hit	
40	<i>Daucus carota</i>	Hit	*
41	<i>Matricaria chamomilla</i>	Hit	*
42	<i>Cymbopogon nardus</i>	Hit	*
D. Disease: female-specific diseases			
1	<i>Foeniculum vulgare</i>	Hit	
2	<i>Imperata cylindrica</i>	Hit	
3	<i>Tamarindus indica</i>	Hit	
4	<i>Pluchea indica</i>	Hit	*
5	<i>Piper retrofractum</i>	Hit	
6	<i>Punica granatum</i>	Hit	
7	<i>Uncaria rhynchophylla</i>	Hit	
8	<i>Zingiber officinale</i>	Hit	
9	<i>Guazuma ulmifolia</i>	Hit	*
10	<i>Nigella sativa</i>	Hit	
11	<i>Terminalia bellirica</i>	Hit	
12	<i>Baeckea frutescens</i>	Hit	
13	<i>Phaseolus radiatus</i>	Hit	
14	<i>Amomum compactum</i>	Hit	*
15	<i>Sauropus androgynus</i>	Hit	
16	<i>Usnea misaminensis</i>	Hit	
17	<i>Cinnamomum burmannii</i>	Hit	
18	<i>Melaleuca leucadendra</i>	Hit	
19	<i>Parameria laevigata</i>	Hit	
20	<i>Parkia roxburghii</i>	Hit	
21	<i>Piper cubeba</i>	Hit	
22	<i>Kaempferia galanga</i>	Hit	

TABLE 4: Continued.

Number	Plants name	Hit-miss status
23	<i>Coriandrum sativum</i>	Hit
24	<i>Kaempferia angustifolia</i>	Hit
25	<i>Curcuma longa</i>	Hit
26	<i>Zingiber aromaticum</i>	Hit
27	<i>Languas galanga</i>	Hit
28	<i>Galla lusitania</i>	Hit
29	<i>Quercus lusitania</i>	Hit
30	<i>Hydrocotyle asiatica</i>	Hit
31	<i>Areca catechu</i>	Hit
32	<i>Lepiniopsis ternatensis</i>	Hit
33	<i>Helicteres isora</i>	Hit *
34	<i>Piper betle</i>	Hit
35	<i>Elephantopus scaber</i>	Hit *
36	<i>Kaempferia pandurata</i>	Hit
37	<i>Curcuma xanthorrhiza</i>	Hit
38	<i>Sesbania grandiflora</i>	Hit
E. Disease: the heart and blood vessels		
1	<i>Allium sativum</i>	Hit
2	<i>Curcuma longa</i>	Hit *
3	<i>Morinda citrifolia</i>	Hit *
4	<i>Homalomena occulta</i>	Hit *
5	<i>Hydrocotyle asiatica</i>	Hit
6	<i>Alstonia scholaris</i>	Hit *
7	<i>Syzygium polyanthum</i>	Miss *
8	<i>Andrographis paniculata</i>	Hit *
9	<i>Apium graveolens</i>	Miss
10	<i>Imperata cylindrica</i>	Hit
F. Disease: male-specific diseases		
1	<i>Cucurbita pepo</i>	Miss
2	<i>Serenoa repens</i>	Miss
3	<i>Baeckea frutescens</i>	Hit
4	<i>Phaseolus radiatus</i>	Hit
5	<i>Curcuma longa</i>	Hit
6	<i>Elephantopus scaber</i>	Hit
G. Disease: muscle and bone		
1	<i>Foeniculum vulgare</i>	Hit
2	<i>Clausena anisum-olens</i>	Hit *
3	<i>Zingiber purpureum</i>	Hit
4	<i>Allium sativum</i>	Hit
5	<i>Strychnos ligustrina</i>	Hit
6	<i>Tinospora tuberculata</i>	Hit *
7	<i>Piper retrofractum</i>	Hit
8	<i>Syzygium aromaticum</i>	Hit
9	<i>Cola nitida</i>	Hit *
10	<i>Ginkgo biloba</i>	Hit *
11	<i>Panax ginseng</i>	Hit
12	<i>Equisetum debile</i>	Hit *
13	<i>Zingiber officinale</i>	Hit

TABLE 4: Continued.

Number	Plants name	Hit-miss status
14	<i>Ganoderma lucidum</i>	Hit
15	<i>Nigella sativa</i>	Hit
16	<i>Terminalia bellirica</i>	Hit *
17	<i>Baeckea frutescens</i>	Hit *
18	<i>Amomum compactum</i>	Hit
19	<i>Cinnamomum burmannii</i>	Hit
20	<i>Melaleuca leucadendra</i>	Hit
21	<i>Parameria laevigata</i>	Hit *
22	<i>Psophocarpus tetragonolobus</i>	Hit *
23	<i>Parkia roxburghii</i>	Hit
24	<i>Piper cubeba</i>	Hit *
25	<i>Kaempferia galanga</i>	Hit
26	<i>Coriandrum sativum</i>	Hit
27	<i>Cola acuminata</i>	Hit
28	<i>Coffea arabica</i>	Hit
29	<i>Orthosiphon stamineus</i>	Hit
30	<i>Curcuma longa</i>	Hit
31	<i>Piper nigrum</i>	Hit
32	<i>Alpinia galanga</i>	Hit
33	<i>Vitex trifolia</i>	Hit
34	<i>Zingiber amaricans</i>	Hit *
35	<i>Zingiber zerumbet</i>	Hit
36	<i>Zingiber aromaticum</i>	Hit
37	<i>Languas galanga</i>	Hit
38	<i>Massoia aromatica</i>	Hit
39	<i>Morinda citrifolia</i>	Hit
40	<i>Carum copticum</i>	Hit *
41	<i>Panax pseudoginseng</i>	Hit *
42	<i>Oryza sativa</i>	Hit
43	<i>Myristica fragrans</i>	Hit
44	<i>Pandanus amaryllifolius</i>	Hit
45	<i>Eurycoma longifolia</i>	Hit
46	<i>Hydrocotyle asiatica</i>	Hit
47	<i>Areca catechu</i>	Hit *
48	<i>Mentha arvensis</i>	Hit *
49	<i>Lepiniopsis ternatensis</i>	Hit
50	<i>Pimpinella pruatjan</i>	Hit
51	<i>Andrographis paniculata</i>	Hit
52	<i>Blumea balsamifera</i>	Hit
53	<i>Cymbopogon nardus</i>	Hit
54	<i>Sida rhombifolia</i>	Hit
55	<i>Cinnamomum sintoc</i>	Hit
56	<i>Piper betle</i>	Hit *
57	<i>Talinum paniculatum</i>	Hit
58	<i>Elephantopus scaber</i>	Hit
59	<i>Cyperus rotundus</i>	Hit
60	<i>Curcuma aeruginosa</i>	Hit
61	<i>Kaempferia pandurata</i>	Hit *

TABLE 4: Continued.

Number	Plants name	Hit-miss status
62	<i>Curcuma xanthorrhiza</i>	Hit
63	<i>Tribulus terrestris</i>	Hit
64	<i>Corydalis yanhusuo</i>	Hit
65	<i>Pausinystalia yohimbe</i>	Hit
<i>H. Disease: nutritional and metabolic diseases</i>		
1	<i>Foeniculum vulgare</i>	Hit
2	<i>Glycyrrhiza uralensis</i>	Hit
3	<i>Zingiber purpureum</i>	Hit
4	<i>Allium sativum</i>	Hit
5	<i>Tinospora tuberculata</i>	Hit
6	<i>Pandanus conoideus</i>	Hit
7	<i>Syzygium aromaticum</i>	Hit
8	<i>Punica granatum</i>	Hit
9	<i>Zingiber officinale</i>	Hit
10	<i>Guazuma ulmifolia</i>	Hit
11	<i>Nigella sativa</i>	Hit
12	<i>Amomum compactum</i>	Hit *
13	<i>Cinnamomum burmannii</i>	Hit
14	<i>Parameria laevigata</i>	Hit
15	<i>Caesalpinia sappan</i>	Hit
16	<i>Soya max</i>	Hit *
17	<i>Cocos nucifera</i>	Hit
18	<i>Rheum tanguticum</i>	Hit
19	<i>Piper cubeba</i>	Hit *
20	<i>Murraya paniculata</i>	Hit
21	<i>Kaempferia galanga</i>	Hit *
22	<i>Coffea arabica</i>	Hit *
23	<i>Orthosiphon stamineus</i>	Hit
24	<i>Curcuma longa</i>	Hit
25	<i>Piper nigrum</i>	Hit *
26	<i>Zingiber aromaticum</i>	Hit
27	<i>Aloe vera</i>	Hit
28	<i>Phaleria papuana</i>	Hit
29	<i>Galla lusitania</i>	Hit
30	<i>Quercus lusitania</i>	Hit
31	<i>Morinda citrifolia</i>	Hit
32	<i>Myristica fragrans</i>	Hit *
33	<i>Momordica charantia</i>	Hit
34	<i>Areca catechu</i>	Hit
35	<i>Lepiniopsis ternatensis</i>	Hit
36	<i>Alstonia scholaris</i>	Hit
37	<i>Hibiscus sabdariffa</i>	Hit
38	<i>Laminaria japonica</i>	Hit
39	<i>Syzygium polyanthum</i>	Hit
40	<i>Andrographis paniculata</i>	Hit
41	<i>Sindora sumatrana</i>	Hit *
42	<i>Cassia angustifolia</i>	Hit
43	<i>Woodfordia floribunda</i>	Hit

TABLE 4: Continued.

Number	Plants name	Hit-miss status
44	<i>Piper betle</i>	Hit
45	<i>Spirulina</i>	Hit
46	<i>Stevia rebaudiana</i>	Hit
47	<i>Theae sinensis</i>	Hit
48	<i>Sonchus arvensis</i>	Hit
49	<i>Curcuma heyneana</i>	Hit
50	<i>Curcuma aeruginosa</i>	Hit
51	<i>Kaempferia pandurata</i>	Hit *
52	<i>Curcuma xanthorrhiza</i>	Hit
53	<i>Curcuma zedoaria</i>	Hit *
54	<i>Olea europaea</i>	Hit
<i>I. Disease respiratory diseases</i>		
1	<i>Foeniculum vulgare</i>	Hit
2	<i>Clausena anisum-olens</i>	Hit
3	<i>Glycyrrhiza uralensis</i>	Hit
4	<i>Zingiber purpureum</i>	Hit
5	<i>Piper retrofractum</i>	Hit *
6	<i>Syzygium aromaticum</i>	Hit
7	<i>Gaultheria punctata</i>	Hit
8	<i>Panax ginseng</i>	Hit
9	<i>Equisetum debile</i>	Hit *
10	<i>Zingiber officinale</i>	Hit
11	<i>Citrus aurantium</i>	Hit *
12	<i>Nigella sativa</i>	Hit *
13	<i>Amomum compactum</i>	Hit
14	<i>Cinnamomum burmannii</i>	Hit
15	<i>Melaleuca leucadendra</i>	Hit
16	<i>Parkia roxburghii</i>	Hit
17	<i>Cocos nucifera</i>	Hit
18	<i>Piper cubeba</i>	Hit
19	<i>Kaempferia galanga</i>	Hit
20	<i>Coriandrum sativum</i>	Hit
21	<i>Curcuma longa</i>	Hit
22	<i>Piper nigrum</i>	Hit
23	<i>Zingiber aromaticum</i>	Hit
24	<i>Languas galanga</i>	Hit
25	<i>Mentha piperita</i>	Hit
26	<i>Oryza sativa</i>	Hit *
27	<i>Myristica fragrans</i>	Hit
28	<i>Pandanus amaryllifolius</i>	Hit *
29	<i>Hydrocotyle asiatica</i>	Hit *
30	<i>Mentha arvensis</i>	Hit
31	<i>Lepiniopsis ternatensis</i>	Hit
32	<i>Helicteres isora</i>	Hit
33	<i>Blumea balsamifera</i>	Hit
34	<i>Cymbopogon nardus</i>	Hit
35	<i>Piper betle</i>	Hit
36	<i>Curcuma xanthorrhiza</i>	Hit

TABLE 4: Continued.

Number	Plants name	Hit-miss status
37	<i>Salix alba</i>	Hit *
38	<i>Matricaria chamomilla</i>	Miss *
<i>J. Disease: skin and connective tissue</i>		
1	<i>Strychnos ligustrina</i>	Hit
2	<i>Merremia mammosa</i>	Hit *
3	<i>Piper retrofractum</i>	Hit *
4	<i>Santalum album</i>	Hit
5	<i>Zingiber officinale</i>	Hit *
6	<i>Citrus aurantium</i>	Hit
7	<i>Citrus hystrix</i>	Hit
8	<i>Cassia siamea</i>	Hit
9	<i>Cocos nucifera</i>	Hit
10	<i>Trigonella foenum-graecum</i>	Hit
11	<i>Orthosiphon stamineus</i>	Hit
12	<i>Curcuma longa</i>	Hit
13	<i>Vetiveria zizanioides</i>	Hit
14	<i>Aloe vera</i>	Hit
15	<i>Rosa chinensis</i>	Hit
16	<i>Jasminum sambac</i>	Hit
17	<i>Phyllanthus urinaria</i>	Hit
18	<i>Mentha piperita</i>	Hit
19	<i>Oryza sativa</i>	Hit
20	<i>Myristica fragrans</i>	Hit *
21	<i>Hydrocotyle asiatica</i>	Hit
22	<i>Lepiniopsis ternatensis</i>	Hit
23	<i>Alstonia scholaris</i>	Hit
24	<i>Andrographis paniculata</i>	Hit
25	<i>Cymbopogon nardus</i>	Hit
26	<i>Piper betle</i>	Hit
27	<i>Theae sinensis</i>	Hit
28	<i>Curcuma heyneana</i>	Hit
29	<i>Kaempferia pandurata</i>	Hit *
30	<i>Curcuma xanthorrhiza</i>	Hit
31	<i>Melaleuca leucadendra</i>	Hit
32	<i>Matricaria chamomilla</i>	Miss *
<i>K. Disease: the urinary system</i>		
1	<i>Foeniculum vulgare</i>	Hit *
2	<i>Imperata cylindrica</i>	Hit *
3	<i>Strychnos ligustrina</i>	Hit *
4	<i>Plantago major</i>	Hit
5	<i>Zingiber officinale</i>	Hit *
6	<i>Cinnamomum burmannii</i>	Hit *
7	<i>Strobilanthes crispus</i>	Hit
8	<i>Kaempferia galanga</i>	Hit *
9	<i>Orthosiphon stamineus</i>	Hit
10	<i>Phyllanthus urinaria</i>	Hit
11	<i>Blumea balsamifera</i>	Hit *
12	<i>Sonchus arvensis</i>	Hit
13	<i>Curcuma xanthorrhiza</i>	Hit

* indicates that plant will not assigned if we use matching score >0.7.

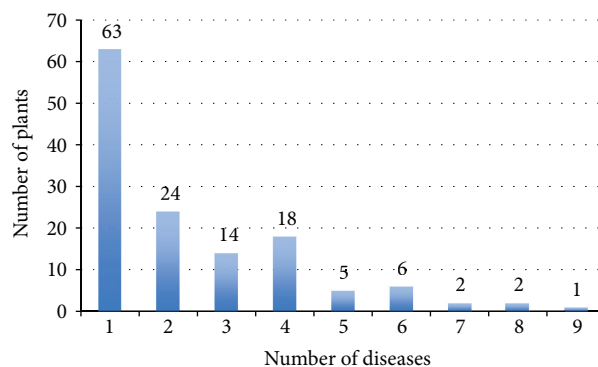


FIGURE 6: Distribution of 135 plants assigned based on 0.7% dataset with respect to the number of diseases they are assigned to.

for any type of application. It ensures coverage and performs robustly in case of random addition, removal, and rearrangement of edges in protein-protein interaction (PPI) networks [14]. While applying DPCLUSO, the parameter values of density and cluster property that we used in this experiment are 0.9 and 0.5, respectively [15]. Table 3 shows the summary of clustering result by DPCLUSO. Because clusters consisting of two Jamu formulas are trivial clusters, for the next steps we only use clusters each of which consists of 3 or more Jamu formulas. The number of total clusters increases along with the larger dataset, although the threshold correlation between Jamu pairs decreases. We evaluated the clustering result using matching score to determine dominant disease for every cluster (step 3 in Figure 1). Matching score of a cluster is the ratio of the highest number of Jamu associated with the same disease to the total number of Jamu in the cluster. Thus matching score is a measure to indicate how strongly a disease is associated to a cluster. Figure 4 shows the distribution of the clusters with respect to matching score from three datasets. All datasets have the highest frequency of clusters at matching score >0.9 and overall most of the clusters have higher matching score, which means most of the DPCLUSO generated clusters can be confidently related to a dominant disease. Furthermore the number of clusters with matching score >0.9 is remarkably larger compared to the same in other ranges of matching score in case of the 0.3% dataset (Figure 4(c)). If we compare the ratio of frequency of clusters at matching score >0.9 for every dataset, the 0.3% dataset has the highest ratio with 40.84% (of 453), compared to 29.67% (of 873) and 21.91% (of 1296), in case of 0.5% and 0.7% datasets, respectively. Thus, the most reliable species to disease relations can be predicted at matching score >0.9 corresponding to the clusters generated from 0.3% dataset.

Figure 5(a) shows the success rate for all 3 datasets with respect to threshold matching scores. Success rate is defined as the ratio of the number of clusters with matching score larger than the threshold to the total number of clusters. As expected it tends to produce lower success rate if we decrease correlation value to create the datasets. However more clusters are generated and more information can be extracted when we lower the threshold correlation value. The success rate increases rapidly as the matching score decreases

TABLE 5: Relation between disease classes in NCBI and efficacy classes reported by Afendi et al. [6].

Class of disease	Ref.	Efficacy class
D1 Blood and lymph diseases	NCBI	E7 Pain/inflammation (PIN)
D2 Cancers	NCBI	E7 Pain/inflammation (PIN)
D3 The digestive system	NCBI	E4 Gastrointestinal disorders (GST) E7 Pain/inflammation (PIN)
D4 Ear, nose, and throat	NCBI	E7 Pain/inflammation (PIN)
D5 Diseases of the eye	NCBI	E7 Pain/inflammation (PIN)
D6 Female-specific diseases	NCBI	E5 Female reproductive organ problems (FML)
D7 Glands and hormones	NCBI	E7 Pain/inflammation (PIN)
D8 The heart and blood vessels	NCBI	E7 Pain/inflammation (PIN)
D9 Diseases of the immune system	NCBI	E7 Pain/inflammation (PIN)
D10 Male-specific diseases	NCBI	E6 Musculoskeletal and connective tissue disorders (MSC)
D11 Muscle and bone	NCBI	E6 Musculoskeletal and connective tissue disorders (MSC)
D12 Neonatal diseases	NCBI	E7 Pain/inflammation (PIN)
D13 The nervous system	NCBI	E7 Pain/inflammation (PIN)
D14 Nutritional and metabolic diseases	NCBI	E2 Disorders of appetite (DOA) E4 Gastrointestinal disorders (GST)
D15 Respiratory diseases	NCBI	E8 Respiratory disease (RSP) E7 Pain/inflammation (PIN)
D16 Skin and connective tissue	NCBI	E9 Wounds and skin infections (WND)
D17 The urinary system	*	E1 Urinary related problems (URI)
D18 Mental and behavioural disorders	*	E3 Disorders of mood and behavior (DMB)

from 0.9 to 0.6 and after that the slope of increase of success rate decreases. Therefore in this study we empirically decide 0.6 as the threshold matching score to predict plant-disease relations.

3.3. Assignment of Plants to Disease. By using DPCLUSO resulting clusters, we assigned plants to classes of disease. Based on a threshold matching score we assigned dominant disease to a cluster. Then we assign a plant to a cluster by way of analyzing the ingredients of the Jamu formulas belonging to that cluster and determining the highest frequency plant, that is, the plant that is used for maximum number Jamu belonging to that cluster (step 4 in Figure 1). Thus we assign a disease and a plant to each cluster having matching score greater than a threshold. Our hypothesis is that the disease and the plant assigned to the same cluster are related.

The total number of assigned plants depends on matching score value. Figure 5(b) shows the number of predicted plants that can be assigned to diseases in the context of matching score. With higher matching score value, the number of predicted plants assigned to classes of disease is supposed to remain similar or decrease but the reliability of prediction increases. In Figure 5(b) a sudden change in the number of predicted plants is seen at matching score 0.6 which we consider as empirical threshold in this work. Based on the 0.7% dataset, the largest number of plants (135 plants, Table 4) was assigned to diseases. There are 63 plants assigned to only one class of disease, whereas the other 72 plants are assigned to at least two or more classes of disease (Figure 6).

3.4. Evaluation of the Supervised Clustering Based on DPCLUSO. We used previously published results [6] as gold standard to evaluate our results. The previous study assigned plants to 9 kinds of efficacy whereas we assigned the plants to 18 disease classes (16 from NCBI and 2 additional classes). For the sake of evaluation we got done a mapping of the 18 disease classes to 9 efficacy classes by a professional doctor, which is shown in Table 5. Table 6 shows the prediction result of plant-disease relations for all 3 datasets, corresponding to clusters with matching score greater than 0.6. Table 6 also shows corresponding efficacy, the number of assigned plants, number of correctly predicted plants, and true positive rates (TPR), respectively.

We determined TPR corresponding to a disease/efficacy class by calculating the ratio of the number of correct prediction to the number of all predictions. When a disease corresponds to more than one kind of efficacy, the highest TPR can be considered the TPR for the corresponding disease. For all 3 datasets the TPR corresponding to each disease is roughly 90% or more. The 0.3% dataset consists of Jamu pairs with higher correlation values and based on this dataset 117 plants are assigned to 14 disease classes. The 0.7% dataset contains more Jamu pairs and assigned plants to 11 disease classes, one less disease class compared to 0.5% dataset. The two disease classes covered by 0.3% dataset but not covered by 0.5% and 0.7% datasets are the nervous system (D13) and disease of the immune system (D9). The only disease class covered by 0.3% and 0.5% datasets but not covered by 0.7% dataset is mental and behavioural disorders (D18). The larger dataset network tends to have

TABLE 6: The prediction result of plant-disease relations using matching score >0.6.

Class of disease	Corresponding efficacy	0.7% dataset			0.5% dataset			0.3% dataset		
		Number of assigned plants	Correct prediction	True positive rate	Number of assigned plants	Correct prediction	True positive rate	Number of assigned plants	Correct prediction	True positive rate
D1	E7	26	22	0.85	24	20	0.83	24	20	0.83
D2	E7	1	1	1.00	5	5	1.00	1	1	1.00
D3	E4	42	42	1.00	33	33	1.00	28	28	1.00
	E7		38	0.90		30	0.91		25	0.89
D4	E7	0	0	—	0	0	—	0	0	—
D5	E7	0	0	—	0	0	—	0	0	—
D6	E5	38	38	1.00	37	37	1.00	32	32	1.00
D7	E7	0	0	—	0	0	—	0	0	—
D8	E7	10	8	0.80	8	7	0.88	6	5	0.83
D9	E7	0	0	—	0	0	—	1	1	1.00
D10	E6	6	4	0.67	2	0	—	3	1	0.33
D11	E6	65	65	1.00	71	71	1.00	60	60	1.00
D12	E7	0	0	—	0	0	—	0	0	—
D13	E7	0	0	—	0	0	—	5	5	1.00
D14	E2	54	44	0.81	45	36	0.80	35	26	0.74
	E4		54	1.00		45	1.00		35	1.00
D15	E7	38	37	0.97	34	34	1.00	33	33	1.00
	E8		31	0.82		30	0.88		29	0.88
D16	E9	32	31	0.97	32	32	1.00	27	27	1.00
D17	E1	13	13	1.00	9	9	1.00	8	8	1.00
D18	E3	0	0	—	5	5	1.00	4	4	1.00
Total assigned plants			135		129			117		

lower coverage of disease classes. The number of Jamu pairs, that is, the number of edges in the network, affect the number of DPCLUSO resulting clusters and number of Jamu formulas per cluster. As a consequence, for the larger dataset networks, the success rate becomes lower and the coverage of disease classes is lower but prediction of more plant-disease relations can be achieved.

4. Conclusions

This paper introduces a novel method called supervised clustering for analyzing big biological data by integrating network clustering and selection of clusters based on supervised learning. In the present work we applied the method for data mining of Jamu formulas accumulated in KNApSAcK database. Jamu networks were constructed based on correlation similarities between Jamu formulas and then network clustering algorithm DPCLUSO was applied to generate high density Jamu modules. For the analysis of the next steps potential clusters were selected by supervised learning. The successful clusters containing several Jamu related to the same disease might be useful for finding main ingredient plant for that disease and the lower matching score value clusters will be associated with varying plants

which might be supporting ingredients. By applying the proposed method important plants from Jamu formulas for every classes of disease were determined. The plant to disease relations predicted by proposed network based method were evaluated in the context of previously published results and were found to produce a TPR of 90%. For the larger dataset networks, success rate and the coverage of disease classes become lower but prediction of more plant-disease relations can be achieved.

Conflict of Interests

The authors declare that there is no financial interest or conflict of interests regarding the publication of this paper.

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References

- [1] R. Verporte, H. K. Kim, and Y. H. Choi, "Plants as source of medicines," in *Medicinal and Aromatic Plants*, R. J. Boger, L. E. Craker, and D. Lange, Eds., chapter 19, pp. 261–273, 2006.
- [2] A. Furnham, "Why do people choose and use complementary therapies?" in *Complementary Medicine: An Objective Appraisal*, E. Ernst, Ed., pp. 71–88, Butterworth-Heinemann, Oxford, UK, 1996.
- [3] E. Ernst, "Herbal medicines put into context," *British Medical Journal*, vol. 327, no. 7420, pp. 881–882, 2003.
- [4] F. M. Afendi, T. Okada, M. Yamazaki et al., "KNAPSAcK family databases: integrated metabolite—plant species databases for multifaceted plant research," *Plant and Cell Physiology*, vol. 53, no. 2, p. e1, 2012.
- [5] F. M. Afendi, N. Ono, Y. Nakamura et al., "Data mining methods for omics and knowledge of crude medicinal plants toward big data biology," *Computational and Structural Biotechnology Journal*, vol. 4, no. 5, Article ID e201301010, 2013.
- [6] F. M. Afendi, L. K. Darusman, A. Hirai et al., "System biology approach for elucidating the relationship between Indonesian herbal plants and the efficacy of Jamu," in *Proceedings of the 10th IEEE International Conference on Data Mining Workshops (ICDMW '10)*, pp. 661–668, Sydney, Australia, December 2010.
- [7] F. M. Afendi, L. K. Darusman, A. H. Morita et al., "Efficacy of Jamu formulations by PLS modeling," *Current Computer-Aided Drug Design*, vol. 9, pp. 46–59, 2013.
- [8] F. M. Afendi, L. K. Darusman, M. Fukuyama, M. Altaf-Ul-Amin, and S. Kanaya, "A bootstrapping approach for investigating the consistency of assignment of plants to Jamu efficacy by PLS-DA model," *Malaysian Journal of Mathematical Sciences*, vol. 6, no. 2, pp. 147–164, 2012.
- [9] W. Winterbach, P. V. Mieghem, M. Reinders, H. Wang, and D. de Ridder, "Topology of molecular interaction networks," *BMC Systems Biology*, vol. 7, article 90, 2013.
- [10] C. Bachmaier, U. Brandes, and F. Schreiber, "Biological network," in *Handbook of Graph Drawing and Visualization*, pp. 621–651, CRC Press, 2013.
- [11] X. Chen, M. Chen, and K. Ning, "BNArray: an R package for constructing gene regulatory networks from microarray data by using Bayesian network," *Bioinformatics*, vol. 22, no. 23, pp. 2952–2954, 2006.
- [12] P. Langfelder and S. Horvath, "WGCNA: an R package for weighted correlation network analysis," *BMC Bioinformatics*, vol. 9, article 559, 2008.
- [13] A. Martin, M. E. Ochagavia, L. C. Rabasa, J. Miranda, J. Fernandez-de-Cossio, and R. Bringas, "BisoGenet: a new tool for gene network building, visualization and analysis," *BMC Bioinformatics*, vol. 11, article 91, 2010.
- [14] M. Altaf-Ul-Amin, M. Wada, and S. Kanaya, "Partitioning a PPI network into overlapping modules constrained by high-density and periphery tracking," *ISRN Biomathematics*, vol. 2012, Article ID 726429, 11 pages, 2012.
- [15] M. Altaf-Ul-Amin, H. Tsuji, K. Kurokawa, H. Asahi, Y. Shinbo, and S. Kanaya, "DPCLUS: a density-periphery based graph clustering software mainly focused on detection of protein complexes in interaction networks," *Journal of Computer Aided Chemistry*, vol. 7, pp. 150–156, 2006.
- [16] S. K. Kachigan, *Multivariate Statistical Analysis: A Conceptual Introduction*, Radius Press, New York, NY, USA, 1991.
- [17] J. L. Rodgers and W. A. Nicewander, "Thirteen ways to look at the correlations coefficient," *The American Statistician*, vol. 42, pp. 59–66, 1995.
- [18] M. Li, J.-E. Chen, J.-X. Wang, B. Hu, and G. Chen, "Modifying the DPCLUS algorithm for identifying protein complexes based on new topological structures," *BMC Bioinformatics*, vol. 9, article 398, 2008.
- [19] World Health Organization, "International Classification of Diseases (ICD) 10," 2010, <http://www.who.int/classifications/icd/en/>.
- [20] National Center for Biotechnology Information, *Genes and Disease*, NCBI, Bethesda, Md, USA, 1998.
- [21] P. Erdos and A. Renyi, "On the evolution of random graph," *Publicationes Mathematicae Debrecen*, vol. 6, pp. 290–297, 1959.
- [22] A.-L. Barabási and R. Albert, "Emergence of scaling in random networks," *Science*, vol. 286, no. 5439, pp. 509–512, 1999.
- [23] A. Vázquez, "Growing network with local rules: preferential attachment, clustering hierarchy, and degree correlations," *Physical Review E—Statistical, Nonlinear, and Soft Matter Physics*, vol. 67, no. 5, Article ID 056104, 15 pages, 2003.
- [24] Max Planck Institut Informatik, "NetworkAnalyzer," 2013, <http://med.bioinf.mpi-inf.mpg.de/netanalyzer/index.php>.



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