

Locus for Malate Secretion in Rice Chromosome 3

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Abstract

Aluminum (Al) toxicity is a major limiting factor of rice production in acid soil. One of the tolerance mechanisms of plant to Al stress is the secretion of malate from plant root. The objective of this research is to identify quantitative trait loci (QTL) in chromosome 3 that controls malate secretion from rice root during the period of Al stress. The research was conducted in laboratory and green house. An F₂ population derived from the cross between rice genotype Hawara Bunar and cultivar IR64 was used in this experiment. Analysis malate secretion was carried out based on enzymatic method. Rice simple sequence repeats from chromosome 3 were as molecular marker. The mapping and QTL analysis was performed using Mapmaker 3 and Mapmaker/QTL. The result showed that the malate secretion trait was normally distributed in the rice F₂ population indicating that the trait is polygenic trait. A QTL for malate secretion was identified in the short arm of rice chromosome 3 located in between markers RM545 and RM517.

Keywords: *Aluminum stress, Chromosome 3, Rice, QTL*

Introduction

In general, aluminum tolerance in plant is genetically controlled, and the diversity of this trait can be found inter and intra plant species, including the member of Gramineae (Aniol and Gustafson 1990). Aluminum tolerance mechanism in plant can be divided into two models (Kochian 1995, Matsumoto 2000). First, internal detoxification mechanism, which is the mechanism of plant cells that are able to detoxify Al in the cell through organic acid chelation (Ma et 1998), vacuolar accumulation, protein detoxification, or reactive oxygen species reduction in the root cells (Ezaki et al 2000). Internal detoxification mechanism can also be achieved through activation of *calmodulin-independent NAD⁺ kinase* (Aniol, 1991).

In the second model, Al is excluded from root tip cells through excluding Al across plasma membrane, increasing rhizosphere pH, producing exudate or secreting organic acid (Taylor 1991, Kochian 1995). Among those external mechanisms, organic acid secretion from root cells, such malate, citrate and oxalate, has been the most accepted mechanism. Organic acid secretion has important role in Al tolerance mechanism in wheat, rye and maize (Ryan et al 1995, Li et al 2002, Pellet et al 1995), however the similar role has not been elucidated in rice.

Several researchers have shown that organic acid secretion closely related to Al tolerance mechanism. However, the main factor that controlled organic acid secretion from root cells during Al stress still unclear. The recent finding showed that over expression of malate transporter gene that isolate from wheat could increase Al tolerance in transgenic tobacco, but the gene could not increase Al tolerance in rice (Sasaki et al 2004).

This paper reported our research in identifying locus for malate secretion trait in rice using an F₂ population derived from a cross between an Indonesian local rice genotype that tolerant to Al and a rice variety IR64 that sensitive to Al. It is expected that the locus can be used to isolate the gene controlling malate secretion in rice.

Materials and Method

Plant Materials

Rice genotype Hawara Bunar (Al-tolerant parent) and cultivar IR64 (Al-sensitive parent), F1 plants and F2 population derived from a cross between Hawara Bunar and IR64 were used in this research.

Nutrient culture technique for aluminum stress treatment

A simple hydroponic technique was used to grow rice seedlings. Seeds were sterilized and soak in destilate water for 24 hours, and followed by germination for 48 hours. The seedlings with homogenous root length were put in sterifoamfloated on sterile minimal nutrient solution (Miftahudin et al. 2004) at pH 4.0 in 15 ml tube. A 15 ppm of Al in the form of $\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$ was administered fro 24 hours. The seedlings were grown at room temperature in growth chamber with 12 hours light photoperiod.

Determiration of malate secretion

Malate secretion was determined from F1 plants, 400 individual F2 population and both parents. Determination of malate secretion followed procedure as described by Delhaize et al. (1993). A 1.35 ml nutrient solution was sampled from each tube and was added with 1.5 ml buffer solution (0.4 M hydrazine and 0.5 M glycine, pH 9) and 0.1 ml 40 mM NAD. The reaction mixtures were thenincubated at room temperaturefor 30-60 min. The absorbance of the mixture was measured using spectrophotometer at $\lambda = 340$ nm (absorbance 1). The mixture was then added with 5 μL Malate Dehydrogenase (5 mg/mL, Sigma, USA), and the absorbance 2 was then measured. The difference between absorbance 1 and 2 indicated the amount of NADH produced and was used to calculate malate content of the solution as the following equation:

$$C = \frac{V \times M}{\epsilon \times d \times v \times 1000} \times \Delta A$$

Notes: C = Malate concentration (mg/l)

V = Volume Total (ml)

v = Sample Volume (ml)

M = Molecular Weight of Malate (g/mol)

d = Cuvette width (cm)

ϵ = Absorption coefficient of NADH at λ 340 nm = 6.3 (l/mmol.cm)

ΔA = the difference between absorbance 1 dan 2

Moelcular marker analysis

The polymorphic SSR markers from rice chromosome 3 were applied to 300 individuals F2 population. The primers from those markers were used to amplify DNA. A 50 ul PCR reaction mixture consist of 100 ng DNA, 100 mM Tris-Cl pH 8.0, 50 mM KCl, 2 mM MgCl_2 , 0.2 mM dNTPs, 0.3-0.5 uM tiap primer, and 1 U Taq DNA polimerase (NEB, USA) was made for each individual plant. PCR process were performed using thermocycler with the following condition: 1 cycle of 94°C for 5 minutes followed by temperature cycles of 94°C (35 second), 55°C (35 second), and 72°C (1 minutes, 45 second) for 35 cycles and finally one cycle of 72°C for 10 minutes . PCR products were analyzed using 2.5 - 3.0% superfine agarose gel electrophoresis in 0.5 x TBE (Tris-Borate-EDTA) buffer.

Genetic mapping and QTL identification

Segregation analysis of molecular marker in the F2 population was performed using *Chi Square* test pada $\alpha = 0.05$. Genetic linkage and QTL analyses used MAPMAKERS/EXP ver 3.0 and MAPMAKERS/QTL ver 1.1, respectively.

Results and Discussion

Malate secretion in parent plants under AI stress

Malate secretion was analyzed from nutrient culture media that has been used for growing F1, F2 and both parent seedling under 72 hours AI stress. The result showed that the malate secretion was higher when the seedlings were AI stressed and the secretion increase as the period of stress increase (Figure 1 and 2). The AI stress at the level of 15 ppm could differentiate malate secretion level from both parents. In rice cv IR64, although malate secretion increase as the increase of stress period, there was no significant difference between malate secretion at 0 and 15 ppm of AI stress along the 72 hour period of stress. Conversely, there was significant difference of malate secretion along the 72 hours stress period between 0 and 15 ppm AI stress in Hawara Bunar. The significant increase of malate secretion occurred when the seedlings were AI-stressed at 15 ppm at 72 hours. Therefore, the AI stress level of 15 ppm for 72 hour stress duration was used for phenotyping the F2 population. The result also showed that the average malate secretion from IR64, Hawara Bunar, F1 and F2 plants were 2.2, 3.8, 3.1 and 4.1 ppm, respectively.

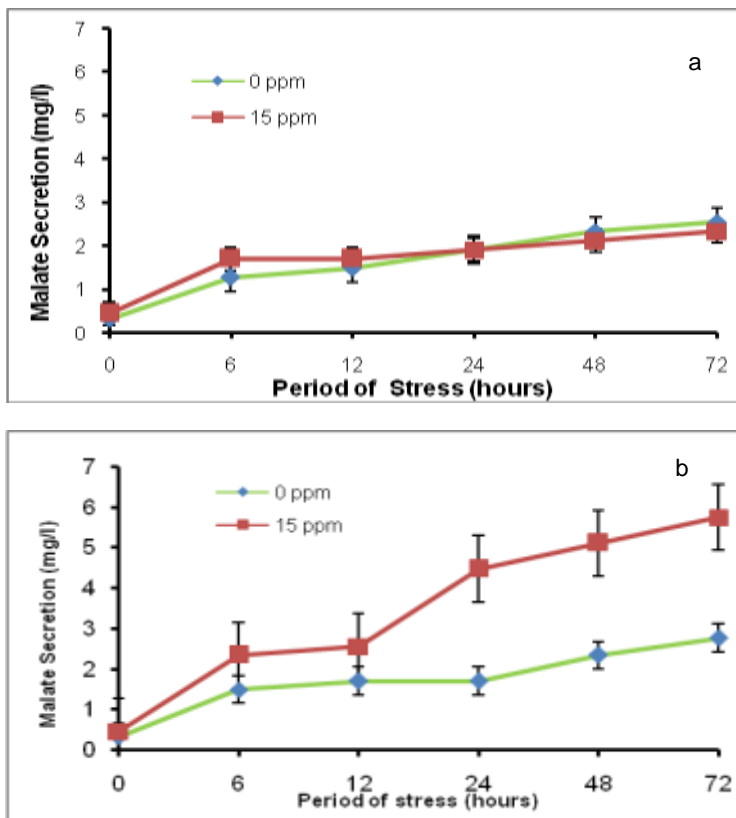


Figure 1. Malate secretion from, rice root cv IR64 (a) and Hawara Bunar (b) along the 72 hour period of AI stress.

Distribution of Malate Secretion in F2 Population

Analysis of malate secretion in 400 individual F2 population showed normal distribution (Figure 2), which indicated that malate secretion character was multigenic controlled. It can be seen from the Figure 2 that the distribution curve rather skewed to the right and the secretion class fallen to higher value than that both parents. This is suggested transgressive segregation phenomenon in this population. The average malate secretion of F2 population was also higher than that of Hawara Bunar.

When the malate secretion was grouped into two class of secretion based on the range of each parent malate secretion, malate secretion in F2 population followed monogenic inheritance. For individual F2 that secreted malate < 3 ppm, it was grouped into AI-sensitive plants, otherwise the plants were grouped into AI-tolerant plants. The Chi square test showed that the segregation fit to 3:1 ratio for AI-tolerant to AI-sensitive plants (Table1).

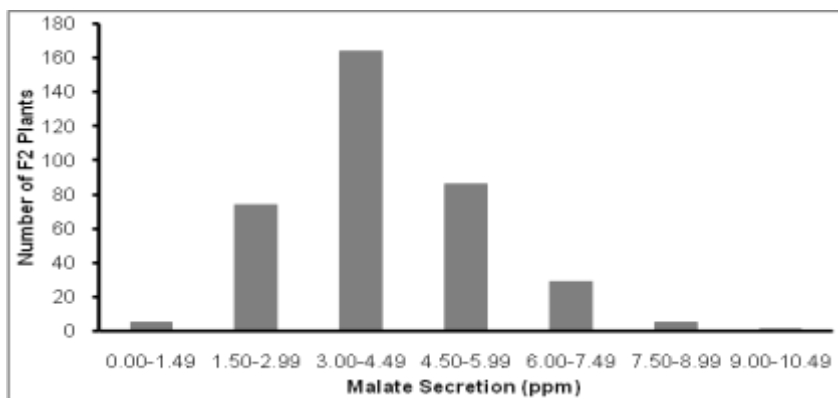


Figure 2. Distribution of malate secretion among individual F2 population.

Table 1. Chi square test for monogenic inheritance pattern of malate secretion in F2 population

Malate Secretion Class (ppm)	Number of Observed Plants	Number of Expected Plants (3:1)	Chi test
< 3.0	79	91	0.15
≥ 3.0	285	273	

Locus for malate secretion in rice chromosome 3

Analysis of genetic factor controlling malate secretion trait in rice was performed into two approaches. First, malate secretion trait was treated as polygenic trait based on the normal distribution of malate secretion in the F2 population, and second, malate secretion trait was treated as monogenic trait. The result showed that among the three chromosome that have been analyzed, which were chromosome 1, 2 and 3, the locus for malate secretion trait was only possibly found on chromosome 3. QTL analysis based on polygenic trait indicated the presence of the QTL in the short arm of chromosome 3 in the region between markers RM517 and RM545. However, the LOD score in that region maximum only 0.98, which was not enough to conclude that the QTL present in that region.

When analysis was performed based on monogenic trait, it was found that a locus for malate secretion trait present in the region with LOD 3.02. Genetic map of the rice chromosome 3 that harbored the locus for malate secretion trait contained five markers i.e. RM569, RM545, RM517, RM251, and RM232 with the total distance 131.8 cM (Figure 3). The locus for malate secretion trait was located in between marker RM545 and RM517 with the exact position at 4 cM

from RM517 toward RM545. The result of this research was similar to Nguyen et al (2001) who found QTL position for Al tolerance loci in the short arm of rice chromosome 3, but the position is rather shifted to other region of the short arm. This difference was due to the different rice background used in both experiments.

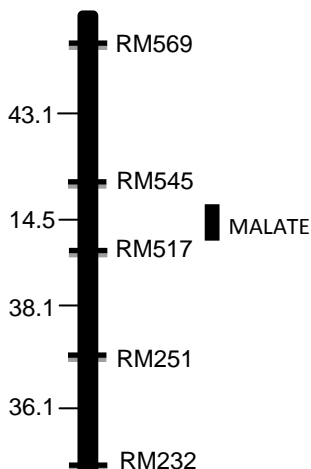


Figure 3. Genetic map of malate secretion locus in rice chromosome 3.

Conclusion

Rice secreted malate during the period of aluminum stress. Al tolerant rice genotype secreted malate more than that of Al-sensitive rice genotype. The secretion increased as the duration of stress increase. There was a locus controlling malate secretion located in the short arm of rice chromosome 3.

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References

- Aniol A (1991) Genetics of acid tolerant plant. In Wright RJ, Baligar VC, Murrmann RP (eds) Plant-Soil Interactions at Low pH. Kluwer Acad Publ, Dordrecht, pp.1007-10017
- Aniol A, Gustafson JP (1990) Genetics of aluminum tolerance in agronomic plants. In Shaw AJ (ed) Heavy Metal Tolerance in Plants: Evolutionary Aspects. CRC Press, Boca Raton, Florida, pp.255-267
- Delhaize E, Ryan PR, Randall PJ (1993b) Aluminum tolerance in wheat (*Triticum aestivum* L.). II. Aluminum-stimulated excretion of malic acid from root apices. Plant Physiol 103:695-702
- Ezaki B, Gardner RC, Ezaki Y, Matsumoto H (2000) Expression of aluminum-induced genes in transgenic Arabidopsis plants can ameliorate aluminum stress and/or oxidative stress. Plant Physiol 122:657-665
- Kochian LV (1995) Cellular mechanisms of aluminum toxicity and resistance in plants. Annu Rev Plant Physiol Plant Mol Biol 46: 237-260
- Li XF, Ma JF, Matsumoto H (2002) Aluminum-induced secretion of both citrate and malate in rye. Plant and Soil 242:235-243
- Ma JF, Hiradate S, Matsumoto H (1998) High aluminum resistance in buckwheat. II. Oxalic acid detoxifies aluminum internally. Plant Physiol 117:753-759
- Matsumoto, H (2000) Cell biology of aluminum toxicity and tolerance in higher plants. Intl Rev Cytol 200:1-46

- Miftahudin, Scoles GJ, Gustafson JP (2004) Development of PCR-based codominant markers flanking the Alt3 gene in rye. *Genome* 47:231-238
- Nguyen VT, Burow MD, Nguyen HT, Le BT, Le TD, Peterson AH (2001) Molecular mapping of genes conferring aluminum tolerance in rice (*Oryza sativa* L.). *Theor Appl Genet* 102:1002-1010
- Pellet DM, Grunes DL, Kochian LV (1995) Organic acid exudation as an aluminum-tolerance mechanism in maize (*Zea mays* L) *Plant* 196:788-795
- Ryan PR, Delhaize E, Randall PJ (1995) Characterization of Al-stimulated efflux of malate from the apices of Al-tolerant wheat roots. *Planta* 196:103-110
- Taylor GJ (1991) Current views of the aluminum stress response: The physiological basis of tolerance. *Curr Topics Plant Biochem Physiol* 10:57-93

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