

## Fragrance Formation in *Aquilaria* spp. Shoot Culture Induced by *Acremonium* sp.

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In this research, fragrance formation in the shoot culture of three clones of *Aquilaria malaccensis* (Ama1, Ama7 and Ama13) and four clones of *A. microcarpa* (Ami5, Ami8, Ami16 and P6) as a response towards infection of four isolates of *Acremonium* sp. (F, G, L, and M) were studied using dual culture methods in three different concentrations of a modified Murashige-Skoog (MSmod) media (50, 75 and 100%). The result indicated that *Acremonium* F and M induced the formation of fragrance. The index of fragrance induced by *Acremonium* F was twice from that of *Acremonium* M, and commonly produced in shoot cultures in MSmod 50%. The index of fragrance in *A. malaccensis* shoot culture was 70% higher than that of *A. microcarpa*. Furthermore, GLC analyses of acetone extracted from fragranced shoots of *A. malaccensis* indicated that only one compound (RT 5.76) might determine the fragrance. Host-microbe compatibility study indicated that the presence of *Acremonium* significantly affected the fitness of the shoots. In contrast, the presence of shoots did not significantly affect the growth rate of *Acremonium*. In general, *Acremonium* F, L and M significantly increased shoot death during the 35 days of interaction.

**Key words:** Fragrance, *Aquilaria*, *Acremonium*, fungal growth response, percentage of dead shoots, fragrance index

Agarwood is produced in trees of *Aquilaria* and *Gyrinops* species. In Indonesia, agarwood is mainly harvested from *A. malaccensis*, *A. microcarpa*, *A. filaria* and *Gyrinops verstepii*. Agarwood is an accumulation of resinaceous substances produced by the tree and deposited in the heartwood. The resin is produced as a part of the tree defense reaction towards mechanical damage or fungal infection. *Acremonium* F has been proved to be a potential agent to induce agar formation in young trees of *Aquilaria crassna*, *A. malaccensis* and *A. microcarpa* (Rahayu *et al.* 1999). It took more than 4 months of incubation for fragrance formation in these trees.

A preliminary study on the dual culture of shoot cultures of *Aquilaria* and *Acremonium* (Rahayu *et al.* 2001) indicated that this method can be utilized to evaluate the capability of agents, both the fungi and the plant hosts, in fragrance formation. About 20 clones of *Aquilaria* have been maintained as shoot cultures, yet the potential of these clones to produce agar substance has not been known. The same thing happened to the *Acremonium* isolated from agarwood. The objective of this study is, therefore, to evaluate the response (in terms of fragrance formation) of three clones of *A. malaccensis* (Ama1, Ama7 and Ama13) and four clones of *A. microcarpa* (Ami5, Ami8, Ami16 and P6) towards four *Acremonium* isolates (F, G, L, M) infection, in three concentrations of modified Murashige Skoog medium (MSmod 100%, 75% and 50%).

### MATERIALS AND METHODS

**Source of Explants and Shoot Cultures.** The stocks of shoot cultures of three clones of *A. malaccensis* (Ama1, Ama7 and Ama13) and four clones of *A. microcarpa* (Ami5, Ami8, Ami16 and P6) of the collection of Biotechnology and Tree Breeding Laboratory, BIOTROP, were used as sources of explants. The shoots were first multiplied by cutting one or two nodes, and the nodes were planted in shoot inducing

media for 2 months. The Murashige Skoog modified media (MSmod) with BAP  $1 \text{ mg L}^{-1}$  (Situmorang 2000) was used as inducing media. The new formed shoots were then elongated for 1 month in MSmod. All cultures were maintained at  $26^{\circ}\text{C}$  with 16 h photoperiods of cool-light of 80 lux. The elongated shoots were served as sources of shoots in dual culture.

**Fungal Isolates and Source of Inoculum.** The stocks of culture of four *Acremonium* sp. isolates (F, G, L, M) of the collection of the Mycology Laboratory, Bogor Agricultural University were used. These isolates were first regenerated in potato sucrose agar in Petri dishes and incubated at room condition. A one-week-old *Acremonium* culture was used as a source of inoculum in a dual culture.

**Fragrance Formation.** Fragrance formation was studied in a dual culture of shoots of a clone of *Aquilaria* sp and an *Acremonium* isolate. The dual culture was made in three concentrations of MSmod, i.e. 100%, 75% and 50%. Firstly, two or three vigorous shoots of *Aquilaria* sp. were planted in MSmod about 2 cm from the edge of the Petri dishes. A small inoculum (0.5 cm diameter) of each culture of *Acremonium* sp. was then planted at a distance of 4.5 cm from the shoots. All treatments were done in 3 replicates. A single culture of either *Acremonium* or shoots was used as control. Prior to determination of fragrance formation, the response of *Acremonium* towards the presence of shoots and the fitness of the shoots in the presence of *Acremonium* were observed. The data obtained were analyzed using univariate analyses of SPSS version 13. The Duncan's multiple range test was used to compare means of treatments. A significant difference was determined at  $\alpha 0.05$ .

The response of *Acremonium* towards the presence of the shoots was stated as the colony radial growth rate (RGR) of the *Acremonium* (Peters *et al.* 1998). The colony growth was observed every two days and stopped when the colony was in contact with the shoots. The response of the shoots was observed weekly until all shoots died or at a maximum of 60 days of interaction and calculated as percentage of dead shoots (PDS).

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The fragrance formation was measured by the organoleptic method and followed by the gas liquid chromatography (GLC) analyses of the fragrant shoots. The organoleptic determination was done by five persons whom have experience in agarwood fragrance and stated as fragrant indexes (FI: 0, no fragrance produced; 1, slightly produced fragrance; 2, moderately produced fragrance; 3, strongly produced fragrance).

**Agar Substance Extraction and Its GLC Analyses.**

When the interaction ended, the fragrance shoots and control shoots were ground in a mortar to make paste. The paste was then extracted in 15 mL of acetone and blended with a magnetic stirrer. The extract was filtrated through Whatman no 1 filter paper which was sprinkled with Na<sub>2</sub>SO<sub>4</sub> anhydrous. The filtrate was concentrated until 1/10 of its volume. The components of agar substances were determined qualitatively using GLC with Carbowax packed column 20 on Chromosorb WAW 80/100 mesh, injector temperature of 23°C, oven column temperature of 195°C and N<sub>2</sub> carrier gas (50 mL/min), air velocity (0.55 kg/cm<sup>2</sup>) and an FID detector. Retention time (Rt) of the components of agar substance from the extracted shoots was compared with that of the natural agarwood. Peaks with similar Rt were considered as similar components of agar substance.

**RESULTS**

**Fragrance Formation.** The fragrance produced (stated as FI) depended on the isolates of *Acremonium*, clones of *Aquilaria* and MSmod concentrations. *Acremonium F* induced fragrance with the highest index (1.73), followed by *Acremonium M* (0.83). Furthermore, *Acremonium G* (0.07) and L (0) were considered not inducing fragrance formation (Fig 1). In general, *A. malaccensis* produced fragrance with index higher than *A. microcarpa*. Amongst the *A. malaccensis* clones, Ama7 produced the highest grade and was significantly different from the two other clones (Fig 2), whilst the fragrance produced by all clones of *A. microcarpa* varied and were significantly different (Fig 2). The FI was also determined by the compatibility of *Acremonium* and the clones. *Acremonium F* and M might induce fragrance production in all clones, while *Acremonium G* induced fragrance only in Ama1 and Ama7.

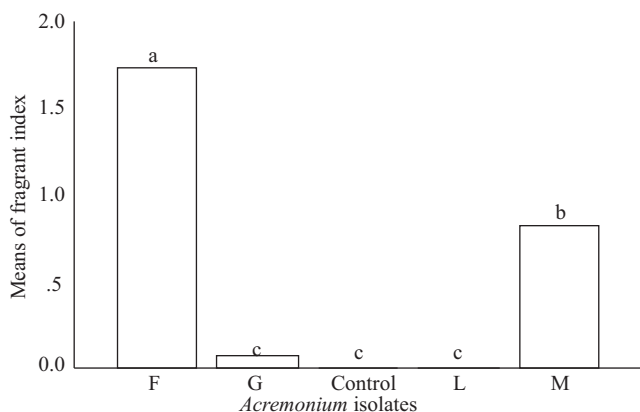


Fig 1 Capability of *Acremonium* isolates to induce fragrance [stated as Fragrance Index (FI). Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05].

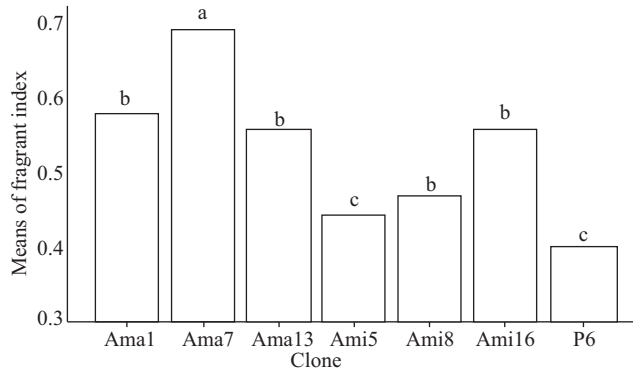


Fig 2 Fragrance produced in various clones of *Aquilaria*. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

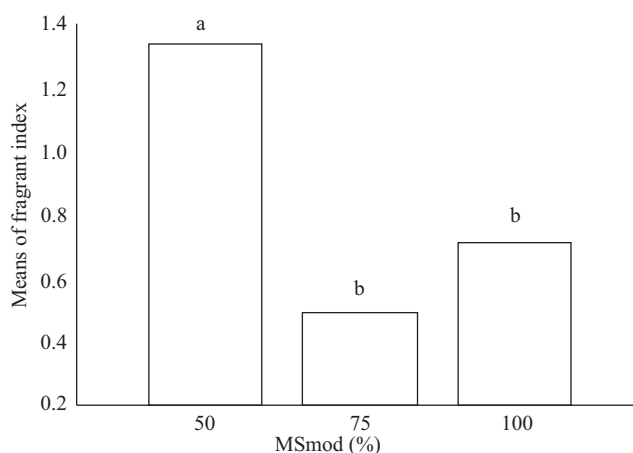


Fig 3 Fragrance induced by *Acremonium F* in shoot planted in various MSmod concentration. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

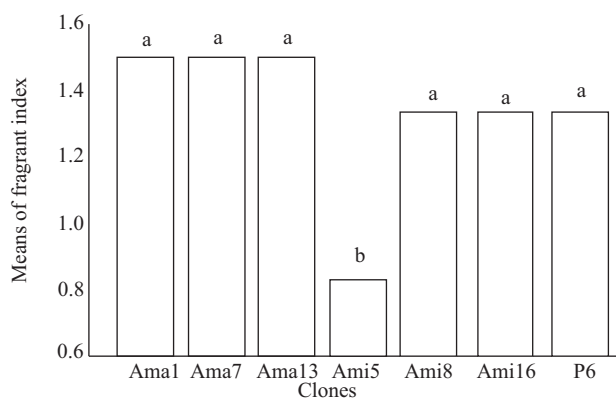


Fig 4 Fragrance produced by various clones as induced by *Acremonium F*. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

Shoots planted in different MSmod concentrations showed FI in significant differences. The shoots planted in MSmod 50% produced the highest FI (1.18), followed by those planted in MSmod 100% (0.65) and MSmod 75% (0.47). This indicated that nutrition limitation enhanced plant stress.

Interaction of the three factors was also observed. As it is too complicated to explain, the result reported here is strictly

on the role of *Acremonium* F in inducing fragrance formation in various clones at various MSmod concentrations. Fragrance grades as induced by *Acremonium* F in all clones are insignificantly different. However, the optimum MSmod concentration for *Acremonium* to induce fragrance is 50% (Fig 3). In this media, all but Ami5 produced an insignificant different FI (Fig 4).

The colony growth was observed for about 3 weeks. In the third week some of the colonies had been in contact with the shoots. During fungal-plant interaction, the colony diameter was neither inhibited nor stimulated (Fig 1). *Acremonium* grew further covering the shoots.

Statistic analyses indicated that the RGR of the colonies of all *Acremonium* isolates were not significantly affected by the presence of the shoots, but by the presence of the media concentration. The RGR of the colonies of *Acremonium* in MSmod 100% and 50% were greater and were significantly different from those of 75% (Table 1). Furthermore, the growth of *Acremonium* G was the slowest and significantly different from the other *Acremonium* isolates (Table 1). The optimal media concentration for each *Acremonium* isolate varied. *Acremonium* F and G grew best on MSmod 50%, but *Acremonium* L on MSmod 100% (Table 1).

Table 1 Radial growth rate\* of *Acremonium* in various concentrations of Msmod

Concentration of MSmod (%)	<i>Acremonium</i>			
	F	G	L	M
50	1.89 <sup>a</sup>	1.26 <sup>b</sup>	1.25 <sup>b</sup>	1.53 <sup>a</sup>
75	1.16 <sup>b</sup>	0.06 <sup>b</sup>	1.25 <sup>b</sup>	1.47 <sup>a</sup>
100	1.35 <sup>a</sup>	0.07 <sup>b</sup>	1.93 <sup>a</sup>	1.89 <sup>a</sup>

\*Stated as rate of radial growth colony increment (mm/day). Data followed by different letters are significantly different by Duncan test at  $\alpha$  0.05.

Shoots started to show browning leaves when in contact with the fungus. However, *Acremonium* L might cause browning leaves prior to contact. Browning leaves usually indicated that the shoots started to die. As a preliminary observation, shoots with brown leaves never recovered. They were considered as dead shoots. Liquid drops were sometimes formed on the surface of the yellowing (Fig 5) or browning leaves.

The PDS was affected significantly in the presence of certain *Acremonium* isolates and MSmod concentrations. *Acremonium* M caused the highest PDS (88.89%), followed by F (69.44%), L (57.54%) and G (23.02%), respectively (Fig 6). In MSmod 100% and 50%, the PDS was greater and significantly different from that of 75% (Fig 7). The PDS was also affected by the interaction between *Acremonium* and MSmod concentrations. *Acremonium* F caused the highest PDS in dual cultures with shoots planted either in MSmod 50% or MSmod 100%. This PDS was not significantly different from the PDS of shoots in dual cultures with M planted in the same concentration as Msmod.

All clones, except for those planted in dual cultures with *Acremonium* F, were not significantly different in their PDS.

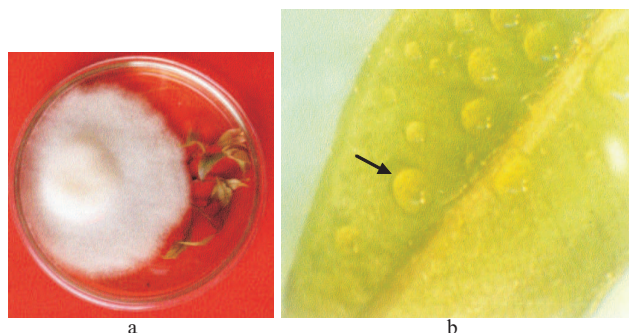


Fig 5 Dual culture of *Acremonium* F and *Aquilaria malaccensis* in 20 days of incubation (a), and liquid drops formed on the lower surface of the leaf of shoot in dual culture (b).

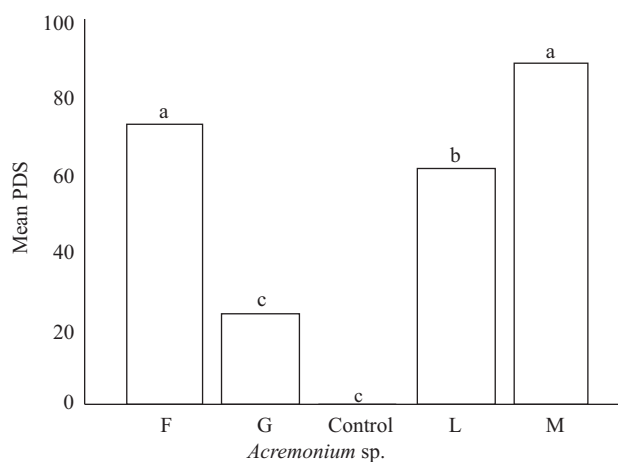


Fig 6 The percentage of death shoot (PDS) caused by various *Acremonium* isolates. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

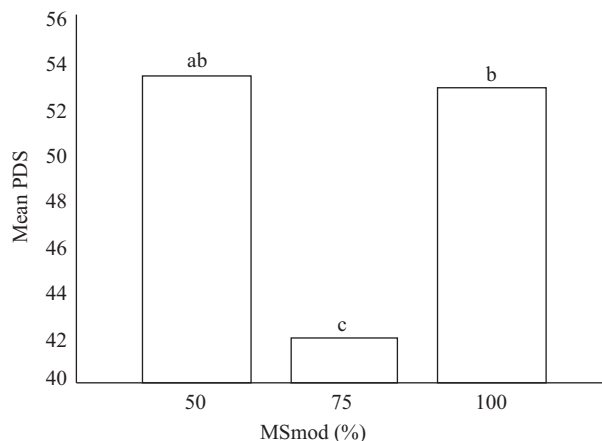


Fig 7 The percentage of death shoot (PDS) in various MSmod concentrations. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

*A. microcarpa* P6 was the most resistant clone, in contrast to Ami 8, which was the most sensitive clone (Fig 8). The PDS of clones also varied significantly when planted in MS mod 50%. *A. microcarpa* Ami 5 was the fittest clone (Fig 9).

**GLC Analyses.** Fourteen components of agarwood substance were detected from natural agarwood. However, the fragrance shoots contained a maximum of 8 components. By comparing to the agarwood substance in shoots which were planted in single cultures, three new components were

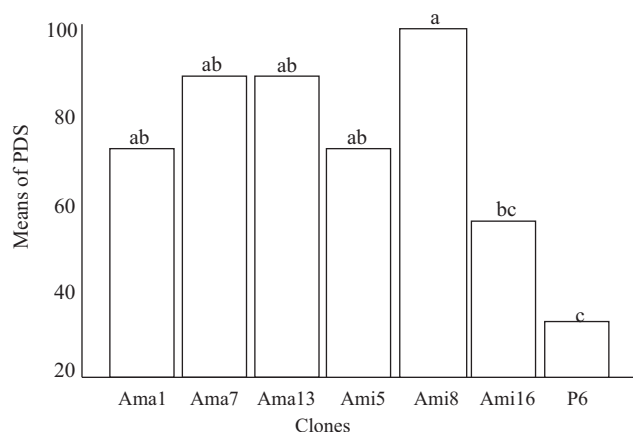


Fig 8 The percentage of death shoot (PDS) of various clones caused by *Acremonium* F. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

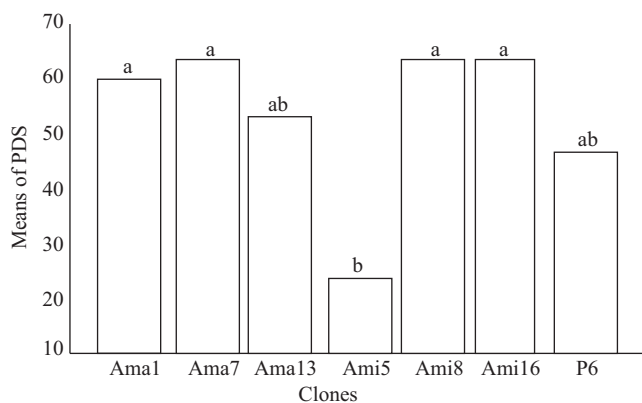


Fig 9 The percentage of death shoot (PDS) of various clones planted in MSmod 50%. Bar with different letters are significantly different by Duncan test at  $\alpha$  0.05.

produced in shoots planted in the dual cultures. These are the substances with Rt 5.76, 22.27, and 27.46. Of those substances, the component with Rt 5.76 was the most frequently formed in dual cultures. This was then considered as the substances that produced fragrance.

## DISCUSSION

**Degree of Fragrance.** A compatible interaction between *Acremonium* and agarwood clones produces specific agarwood fragrance. *Acremonium* F and M showed a wider compatibility in terms of host species ranges rather than did *Acremonium* G and L. Although the fragrance produced among host species or within species was significantly different, the capability of individual shoots to give response to infection should also be counted. Age and Hill (1994) asserted that the response given by the tall fescue due to *Acremonium* infection varied depending on the plant genotype. There are genotypes with high concentration of ergovaline and those with low concentration of ergovaline.

Not all shoots were capable of producing fragrance. The mechanism of response is not clear yet. In the controlled condition, as in the dual culture, the amount of nutrition available might be critical in producing fragrance. Shoots that grew in half the strength of MSmod medium were

capable of producing fragrance stronger than those that grew in full strength of MSmod medium. This indicated that when the plant was stressed, more fragrance was produced. According to Ramawat and Sonie (1999), secondary metabolite was produced when plant was under stress. Therefore, the degree of fragrance might not be a specific reaction to infection, but rather a response to all kinds of stress.

Although it has been stated that nutrition can be a critical factor for fragrance formation, the media for shoot cultures are generally suitable for *Acremonium* growth. On the basis of their RGR, it is known that the behavior of two *Acremonium* isolates, i.e. F and G, is different from the others. Isolates F and G grew best at 50% while the other grew best at 100%. The differences in behavior toward nutrition may be due to the differences in the sources of isolates. *Acremonium* F was obtained from *G.verstegii* (Lombok), whereas the G and L isolates were from *A. malaccensis* (Riau), and M was from unknown agarwood (Papua). Character differences among *Acremonium* isolates had been reported by Carson (1999), who reported isolates of *Acremonium* from Cucurbitaceae. She mentioned that these *Acremonium* might differ in their production of chlamyospores. Some isolates showed prolific production of chlamyospores, while others produced only a small number. Rahayu (2001) also reported that *Acremonium* from agarwood could be differentiated on the basis of their sensitivity to benomil, but their sensitivity to benomil did not depict their RGR in MSmod.

The RGR of *Acremonium* isolates were not influenced by the presence of the shoots. It might be no host secretion that affected the fungal growth. This is in contrast to Sieber *et al.* (1990), who stated that the growth of *Cryptodiaphorthe hytrix* was strongly stimulated by the presence of host callus *Acer macrophyllum*. Water-soluble metabolites produced by the callus may be responsible for this stimulation. Similarly, Lu and Clay (1994) stated that *Atkinsonella* grew faster on their native hosts than on alien hosts. The growth of the *Atkinsonella* isolated from *Danthonia sericea* was stimulated by *D. sericea* calli, whereas the *D. spicata* isolate was inhibited on the third day of growth relative to the controls. Furthermore, it was stated that the dual culture represents an in vitro system for screening compatibility of host-pathogen, in this case clavicipitaceous fungi with both native and alien hosts, according to Peters *et al.* (1998)

The presence of shoots did not affect the RGR of *Acremonium*. However, the presence of *Acremonium* significantly reduced the health of the shoots. The shoots were dead after being in contact with *Acremonium*, except for those that were paired with *Acremonium*. This indicated that *Acremonium* may affect shoot health in a different way.

To initiate an infection, germ-tubes of these *Acremonium* do not produce appressorium. Therefore, the infection may not be due to a mechanical process. According to Pitson *et al.* (1997), *Acremonium* and *Cephalosporium* produced  $\beta$ -glucanase and  $\beta$ -glucosidase that might destroy the polysaccharides of the plant cell walls. These enzymes were inducible. Therefore, these enzymes would only be

produced by the fungus when the fungus was in contact with the plant. This might explain why the vigorousness of the shoots was reduced after being in contact with the fungus. Furthermore, *Cephalosporium graminearum* was reported to produce graminin which might cause the wilting of the leaves of *Triticum aestivum* (Rahman *et al.* 2001). Further research on the ability of these isolates to produce certain types of enzymes and toxins should be conducted.

The pathogenicity of different isolates also varied. Like other phytopathogenic fungi, this phenomenon is quite common. Although M is the most pathogenic isolate, the fragrance produced by the dying shoots is lower than that by *Acremonium* F. The resistance of the Aquilarian clones towards *Acremonium* F also varied and was not correlated with the fragrance produced.

**GLC Analyses.** Fourteen components of agarwood substance were detected from natural agarwood. According to Yuan (1995), about 30 sesquiterpenoids compound have been detected in natural agarwood originated from various *Aquilaria* spp. Among those components, oxo-agarospirol was considered as the components responsible for the fragrance. In the shoots of the dual cultures, only one component with Rt 5.76 out of 8 components detected was considered as the substances that produced fragrance. This compound has not yet identified.

#### ACKNOWLEDGEMENT

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